



Original Scientific Article

**INFLUENCE OF THE SEASON ON THE METABOLIC
PROFILE IN CHIOS SHEEP**

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ABSTRACT

Chios is a breed of sheep selected for milk production, with metabolic features typical for a dairy sheep breed. The energy requirements of pregnant sheep increase in the last weeks of gestation. Metabolic imbalance in the late pregnancy in sheep, usually cause a metabolic disorder known as pregnancy toxemia. Additionally, a pregnant sheep exposed to low environmental temperatures has increased energy demands, due to its adaptation to undesirable environmental conditions. The aim of this study was to compare the metabolic profile of Chios sheep exposed to different environmental conditions. Two groups of ewes were instigated. First group included 8 pregnant ewes with clinical signs of pregnancy toxemia exposed to cold stress during the winter season. The second group included 8 non-pregnant, clinically healthy ewes, that were examined during the non-breeding period, in the spring season. Blood samples were taken and serum concentrations of glucose, beta-hydroxybutyrate (BHBA), total protein, albumin, urea, creatinine, triglyceride and cholesterol, as well as activity of AST and ALP were determined. Pregnant ewes exposed to cold stress had significantly lower levels of glucose and total protein, and significantly higher levels of BHBA, albumin and AST in the serum compared to non-pregnant ewes that were in optimal environmental conditions. There was no significant difference between the serum levels of urea, creatinine, cholesterol, triglycerides and ALP among the groups. In conclusion, low environmental temperature and poor feeding during the winter season caused metabolic distress in pregnant ewes during the early winter season.

Key words: sheep, season, metabolic profile

INTRODUCTION

Ruminants have different carbohydrate metabolism compared to non-ruminant animals. Namely, dietary glucose in sheep and other ruminants is converted into short-chain volatile fatty acids in the rumen, so the main source of

blood glucose is gluconeogenesis in the liver (1), with gluconeogenesis evident even in the fetal sheep liver (2). This also causes blood glucose concentrations to be lower than in non-ruminant animals, and predispose the animal to a negative energy balance (NEB), when energy demands arise. In pregnant ewes, the increased fetal demand is not combined with increased endogenous glucose production, so maternal tissues manifest a relative energy deficit, compared with non-pregnant ewes (3). This energy deficit causes mobilization of fatty acids from the adipose tissue which are utilized in the liver, leading to increased level of ketone bodies, especially beta-hydroxybutyrate (BHBA) (4). This condition could progress into a very severe metabolic disorder - pregnancy toxemia (5), a disease manifested with neural signs due to the

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fact that glucose is the only energy source for the brain (6). There are various factors which have an impact on energy metabolism and contribute to the development of a negative energy balance (NEB). Those are food intake, starvation (7,8), hormonal status (9,10,11,12,13,14), age and gender (15), twin-bearing (16) and cold stress (17). Pregnancy in sheep commonly occurs during the winter season when animals might be exposed to cold stress. Exposure to low environmental temperatures decreases insulin secretion (17). In the condition of a disturbed energy balance, some biochemical parameters in the serum, which are indicators of metabolic disorder and hepatic insufficiency could be changed. These parameters could also be changed due to exposure of animals to extreme environmental temperatures, as was found in cattle (18). Therefore, the aim of this study was to determine possible seasonal variations of biochemical parameters in Chios sheep.

MATERIAL AND METHODS

Multiparous Chios ewes from one flock in Pelagonia, a region in R. of Macedonia, were chosen for the study. One group of ewes (n=8) were chosen during the early winter season (December), when cows were exposed to cold stress, while the other group (n=8) was chosen during the spring season (May). Ewes examined during the winter season were pregnant and manifested clinical signs

of pregnancy toxemia (inappetence, ataxia, lethargy, weight loss) and ewes investigated during the spring season were non-pregnant and clinically healthy. During the winter period, the flock was exposed to environmental low temperature and consumed frozen low quality roughage hay, that caused many lesions in the oral mucosa. According to the data from the flock farm, ewes were not treated for endoparasites. Blood samples were taken from each animal included in study, by venepuncture from *v. jugularis*. Biochemical parameters that were measured in the blood samples were glucose, BHBA, total protein, albumin, urea, creatinine, triglycerides (TG), cholesterol, AST and ALP. Glucose, total protein, albumin, triglyceride, cholesterol were determined by standard "end point" methods according to the manufacturer's instructions Human, Germany. AST, ALP (Human, Germany) and BHBA (Randox, United Kingdom) were measured by kinetic methods according to the manufacturer's instructions, on semiautomatic photometer STAT Fax 3300 (Awareness Technology, Inc, USA).

RESULTS

The obtained results for concentrations of glucose, BHBA, total protein, albumin, urea, creatinine, triglycerides and cholesterol, and for activity of AST and ALP in examined ewes are shown in Table 1.

Table 1. Concentrations (X±SE) of some biochemical parameters in ewes during different seasons. P values indicate the seasonal effect

Parameter	Pregnant ewes during the winter season	Non-pregnant ewes during the spring season	P value
	mean	mean	n.s
glucose (mmol/L)	1.74 ±0.17	2.96±0.14	< 0.01
BHBA (mmol/L)	1.35±0.05	0.69±0.07	< 0.001
total protein (g/L)	57.56±1.89	68.23±1.25	< 0.01
albumin (g/L)	41.30±2.08	32.05±0.91	< 0.01
urea (mmol/L)	5.47±0.44	5.95±0.50	n.s
creatinine (μmol/L)	116.88±5.76	109.82±6.51	n.s
triglycerides (mmol/L)	0.46±0.13	0.27±0.03	n.s
cholesterol (mmol/L)	2.19±0.21	1.57±0.11	n.s
AST (U/L)	191.49±25.71	95.86±7.09	< 0.05
ALP (U/L)	196.55±26.42	128.35±9.09	n.s

According to the results, the group of pregnant ewes during the winter session had significantly lower level of glucose and total protein (1.74 ± 0.17 mmol/L, and 57.56 ± 1.89 g/L, respectively) than non-pregnant ewes during spring session (2.96 ± 0.14 mmol/L, and 68.23 ± 1.25 g/L, respectively) ($p < 0.01$), and significantly higher level of albumin (41.30 ± 2.08 g/L vs. 32.05 ± 0.91 g/L, respectively) ($p < 0.01$). BHBA levels were significantly higher in pregnant ewes during the winter season (1.35 ± 0.05 mmol/L) than in non-pregnant ewes during the spring season (0.69 ± 0.07 mmol/L) ($p < 0.001$), and also this was found in levels of AST (191.49 ± 25.71 U/L in pregnant ewes vs. 95.86 ± 7.09 U/L in non-pregnant ewes) ($p < 0.05$). There were no significant differences in values of urea, creatinine, triglycerides, cholesterol and ALP between two groups.

DISCUSSION

Pregnant ewes have high energy demands due to intensive fetal growth during the last few weeks of pregnancy (14). Pregnant sheep exposed to extremely low environmental temperatures have to increase generation of heat for maintaining body temperature. This could be obtained by intake of a high-energy diet (19). Due to increased energy demands of pregnant ewes, adaptation to environmental conditions could be compromised, especially if cold exposure is accompanied with fasting. Common metabolic disorder in late pregnancy in sheep, especially in sheep with two or more lambs, known as gravidity toxemia is usually caused by metabolic imbalance and environmental stress (5).

Many investigations with sheep exposed to cold (17, 19, 20, 21, 22) have shown that extremely low environmental temperature may cause hyperglycemia, due to decreased secretion of insulin from the pancreas (21). This is opposite to our results, which showed significant hypoglycemia in sheep with pregnancy toxemia during the winter season, when animals were exposed to extremely low environmental temperature and decreased feed intake. Contrary to this, during the spring period animals showed normoglycemia, which is in accordance with the results obtained on sheep exposed to normal environmental conditions (17). There was significant difference ($p < 0.001$) in the glucose level between ewes exposed to cold stress (1.74 ± 0.79 mmol/L) and ewes not exposed to cold stress (2.96 ± 0.44 mmol/L). This finding obviously differs from results of others, probably because the latter included fattening breeds of sheep Suffolk (17, 19, 20), Suffolk-Hampshire-Rambouillet crossbred (21),

Corridale-Suffolk crossbred (22), respectively. This suggests possible significant differences in energy metabolism in dairy and fattening breeds of sheep. In pregnant ewes gluconeogenesis does not increase with increased fetal demand, which may cause energy deficit of maternal tissues. This is not a case in non-pregnant ewes (3). Low concentration of serum glucose in pregnant ewes is probably the consequence of inappropriate gluconeogenesis in the liver from glucogenoplastic precursors, like propionate that originate from rumen fermentation. Food deprivation, which was evident in pregnant ewes during the winter season, causes lack of propionate precursors for glucose production. Biochemical pathways are further complicated with inappropriate beta-oxidation of acetate and butyrate leading to increased production of acetyl-Co-A by hepatocytes. This product is a precursor for acetoacetyl-Co-A. Acetoacetyl-Co-A is a primordial molecule for ketone bodies with high effect on acido-base status. This molecule is converted into a very unstable molecule acetoacetate. Reduction of acetoacetate produces BHBA, while irreversible decarboxylation of acetoacetate produces acetone molecules. Excessive concentration of serum BHBA cannot be utilized by extrahepatic tissue, and therefore the serum level of BHBA remains high. Significantly marked hyperketonemia was noticed in pregnant (1.35 ± 0.27 mmol/L), but not in non-pregnant sheep (0.69 ± 0.21 mmol/L), though it was not as high as found by other authors (24). The difference in BHBA concentrations between pregnant and non-pregnant sheep was significant ($p < 0.001$), due to decreased alimentary energy resources in pregnant sheep. BHBA is an indirect indicator of negative energy balance, and season variation during the late spring period showed lower serum concentration as a result of better energy supply.

Insufficient energy derived by carbohydrate metabolism causes lipolysis, but energy derived from free fatty acids cannot be adequately used in Krebs cycle, because of a lack of oxalacetate, derived from glycogenic precursors. The hepatic capacity for metabolizing of non-esterified fatty acids (NEFA) released from adipose tissue is overloaded. Physiologically, NEFA can be reesterified in triglycerides which are included in the formation of VLDL for secretion in blood. Sheep have a very low capacity for production of VLDL (24), especially breeding and non-breeding sheep compared with sheep in lactation (13), and thereby not capable to produce enough VLDL in condition of compromised lipid metabolism in hepatocytes. In that case, TG can not be removed from hepatocytes, remaining in the hepatocytes

causing hepatic insufficiency. In our study, this was evident by higher serum activity of AST (191.49 ± 117.84 U/L) in pregnant ewes which suffered from severe liver impairment and had decreased liver capacity to maintain gluconeogenesis, compared to non-pregnant ewes (128.35 ± 27.27 U/L) ($p < 0.05$). AST is a liver enzyme that enhances the process of transamination, with oxalacetate, a main metabolite in Krebs cycle, as a final product. As a cytosol enzyme with many isoforms it can easily pass the hepatocyte membrane, so increased serum activity of AST indicates an over loaded hepatocyte activity. Thus, in our study the liver was affected by metabolic distress during the critical period and further complicated by lack of alimentary glycolytic precursors. Homeostatic regulatory mechanisms are disrupted, because of neuro-endocrinological requirements in the period of late gestation and lambing. This critical period in the sheep is further complicated with low environmental temperature and high metabolic energy demand. Variations in environmental conditions during the cold season reflects on the energy metabolism, presented through biochemical parameters. So environmental stress, inappropriate feeding management and inappropriate dehelminthisation decompensate liver function and capacity to survive the critical period. Stress hormones, such as high values of serum cortisol level, further enhance the catabolic processes in the extra mammary tissue and skeletal muscles, affecting the body condition score. It is known that pregnancy toxemia is accompanied with increased cortisol level (24).

Particularly, serum protein fractions pattern could give information about dehydration, plasma volume expansion and hepatic function. Particularly, serum proteinfractions pattern could give information about dehydration, plasma volume expansion and hepatic function.

Another indicator of liver dysfunction are serum proteins levels, especially albumin, which decrease during hepatic insufficiency. In our study, the albumin level was significantly higher during the winter period in pregnant sheep (41.30 ± 9.53 g/L) compared with non-pregnant sheep (32.05 ± 2.73 g/L) ($p < 0.001$), but in fact this was probably pseudo hyperalbuminemia, as the result of dehydration, which is related with changes in particular serum protein fractions (25). Dehydration of sheep appears as a result of the osmotic activity of ketone bodies, because they are very strong acids causing keto-acidotic condition. Lack of water consumption also is one of the main reason for dehydration. Serum concentration of total proteins (68.23 ± 3.76 g/L) showed significantly higher values in non-pregnant sheep during the late spring period, because of regular

alimentary protein precursors supply, compared to underfed pregnant sheep (57.65 ± 5.44 g/L). Status of proteinemia clearly revealed appropriate synthesis and utilization when amino acids precursors are supplied from alimentary recourses. Statistically significant lower concentration of total proteins ($p < 0.001$) in pregnant ewes during the winter period is a result of proteolysis, caused by energy deficit.

There was no significant difference between serum concentrations of urea, creatinine, cholesterol and ALP in pregnant sheep during the winter period (5.47 ± 2.04 mmol/L; 116.88 ± 26.41 μ mol/l; 2.19 ± 1.00 mmol/L, 196.55 ± 121.10 U/L, respectively), and non-pregnant sheep during the spring period (5.95 ± 1.51 mmol/L; 109.82 ± 19.54 μ mol/l; 1.57 ± 0.35 mmol/L; 128.35 ± 27.27 U/L respectively), and these values were all within physiological ranges (26). Also, serum concentrations of triglycerides (0.46 ± 0.60 mmol/L) in pregnant sheep showed mild decrease of values, probably due to reesterification of triglycerides in hepatocytes, but it was not statistically significant compared with non-pregnant sheep (0.27 ± 0.10 mmol/L).

CONCLUSION

Metabolic pathways in pregnant ewes were not able to overcome the negative energy balance during the winter period. On the contrary, the spring period favorably affected the energy status of Chios sheep. During the winter period, the Chios sheep were in an inadequate energy status during late pregnancy and lambing when poor feeding management occurred. Glucose and BHBA concentrations could be reliable indicators of negative energy balance and gravidity toxemia in Chios sheep.

REFERENCES

1. Wang, J., Zhu, X., Chen, C., Li, X., Gao, Y., Li, P., Zhang, Y., Long, M., Wang, Z., Liu, G. (2012). Effect of insulin-like growth factor-1 (IGF-1) on the gluconeogenesis in calf hepatocytes cultured in vitro. *Mol Cell Biochem*, 362, 1-2, 87-91. <http://dx.doi.org/10.1007/s11010-011-1130-9> PMID:22015655
2. Thorn, S.R., Sekar, S.M., Lavezzi, J.R., O'Meara, M.C., Brown, L.D., Hay Jr., W.W., Rozance, P. J. (2012). A physiological increase in insulin suppresses gluconeogenic gene activation in fetal sheep with sustained hypoglycemia. *Am J Physiol Regul Integr Comp Physiol* 303: R861-R869 <http://dx.doi.org/10.1152/ajpregu.00331.2012>

3. Raoofi, A., Jafarian, M., Safi, S., Vatankhah, M. (2013). Fluctuations in energy-related metabolites during the peri-parturition period in Lori-Bakhtiari ewes. *Small Rum Res.* 109, 1, 64-68.
<http://dx.doi.org/10.1016/j.smallrumres.2012.06.012>
4. Moallem, U., Rozov, A., Gootwine, E., Honig, H. (2012). Plasma concentrations of key metabolites and insulin in late-pregnant ewes carrying 1 to 5 fetuses. *JAS*, 90, 1, 318-324.
<http://dx.doi.org/10.2527/jas.2011-3905>
5. Radostits O, Gay C, Hinchcliff K., Constable P. (2006). *Veterinary Medicine - A textbook of the diseases of cattle, horses, sheep, pigs and goats*, 10th Edition. Saunders Ltd.
6. Duehlmeier, R., Fluegge, I., Schwert, B., Ganter, M. (2013). Insulin sensitivity during late gestation in ewes affected by pregnancy toxemia and in ewes with high and low susceptibility to this disorder. *J Vet Intern Med.* 27, 2, 359-366.
<http://dx.doi.org/10.1111/jvim.12035>
PMid:23397990
7. Verbeek, E., Waas, J.R., Oliver, M.H., McLeay, L.M., Ferguson, D.M., Matthews, L.R. (2012). Motivation to obtain a food reward of pregnant ewes in negative energy balance: Behavioural, metabolic and endocrine considerations. *Hormones and Behavior*, 62, 2, 162-172.
<http://dx.doi.org/10.1016/j.yhbeh.2012.06.006>
PMid:22789465
8. Cal-Pereyra, L., Benech, A., González-Monta-a, J.R., Acosta-Dibarrat, J., Da Silva, S., Martínez, A. (2015). Changes in the metabolic profile of pregnant ewes to an acute feed restriction in late gestation. *N Z Vet J.* 63 (3): 141-146.
<http://dx.doi.org/10.1080/00480169.2014.971083>
PMid:25275560
9. Sosa, C., Forcada, F., Meikle, A., Abecia, J.A. (2013). Increase in ovine plasma cortisol at oestrus and its relation with the metabolic status during the sexual cycle in sheep *Biological Rhythm Research*, 44, 3, 445-449.
<http://dx.doi.org/10.1080/09291016.2012.704793>
10. Plested, P.C., Taylor, E., Brindley, D.N., Vernon, R.G. (1987). Interactions of insulin and dexamethasone in the control of pyruvate kinase activity and glucose metabolism in sheep adipose tissue. *Biochem. J.* 247 (2), 459-465.
PMid:3322264 PMCID:PMC1148430
11. Vernon, R.G., Taylor, E. (1988). Insulin, dexamethasone and their interactions in the control of glucose metabolism in adipose tissue from lactating and nonlactating sheep. *Biochem. J.* 256 (2), 509-514.
PMid:3066347 PMCID:PMC1135439
12. Vernon R.G., Finley, E. (1988). Roles of insulin and growth hormone in the adaptations of fatty acid synthesis in white adipose tissue during the lactation cycle in sheep. *Biochem. J.* 256 (3), 873-878.
PMid:2465000 PMCID:PMC1135497
13. Emmison, N., Agius, L. Zammit, V.A. (1991). Regulation of fatty acid metabolism and gluconeogenesis by growth hormone and insulin in sheep hepatocyte cultures - Effects of lactation and pregnancy. *Biochem. J.* 274 (1), 21-26.
PMid:2001235 PMCID:PMC1149914
14. Regnault, T.R.H., Oddy, H.V., Nancarrow, C., Sriskandarajah, N. Scaramuzzi, R.J. (2004). Glucose-stimulated insulin response in pregnant sheep following acute suppression of plasma non-esterified fatty acid concentrations. *Reprod Biol Endocrinol*, 2:64
PMid:15352999 PMCID:PMC519029
<http://dx.doi.org/10.1186/1477-7827-2-64>
15. Kiran, S., Bhutta, A.M., Khan, B.A., Durrani, S., Ali, M., Ali, M., Iqbal, F. (2012). Effect of age and gender on some blood biochemical parameters of apparently healthy small ruminants from Southern Punjab in Pakistan. *Asian Pac J Trop Biomed.* 2 (4): 304-306.
PMCID: PMC1283008
[http://dx.doi.org/10.1016/S2221-1691\(12\)60028-8](http://dx.doi.org/10.1016/S2221-1691(12)60028-8)
16. Rumball, C.W.H., Harding, J.E., Oliver, M.H., Bloomfield, F. H. (2008). Effects of twin pregnancy and periconceptional undernutrition on maternal metabolism, fetal growth and glucose-insulin axis function in ovine pregnancy. *J Physiol* 586 (5), 1399-1411.
<http://dx.doi.org/10.1113/jphysiol.2007.144071>
17. Sasaki, Y., Takahashi, H. (1980). Insulin Secretion in Sheep Exposed to Cold. *J. Physiol.*, 306, 323-335.
<http://dx.doi.org/10.1113/jphysiol.1980.sp013399>
18. Ulcar I., Celeska I. (2010). Seasonal variations of serum biochemical parameters in dairy cows. *Abstracts of Days of Veterinary Medicine 2010*, 28-30 October 2010, Ohrid. *Mac. Vet. Rev.* 33 (2), 30
19. Sano, H., Matsunobu, S., Abe, T., Terashima, Y. (1992). Combined Effects of Diet and Cold Exposure on Insulin Responsiveness to Glucose and Tissue Responsiveness to Insulin in Sheep. *J. Anim. Sci.* 70 (11), 3514-3520.
PMid:1459914
20. Sasaki, Y., Takahashi, H. (1983). Insulin response to secretagogues in sheep exposed to cold. *J Physiol.* 334: 155-167.
<http://dx.doi.org/10.1113/jphysiol.1983.sp014486>
21. Terashima, Y., Tucker, R.E., Deetz, L.E., Degregorio, R.M., Muntifering, R.B., Mitchell, G.E. Jr. (1982) Plasma magnesium levels as influenced by cold exposure in fed or fasted sheep. *J Nutr.* 112(10): 1914-1920.
PMid:6750054
22. Sano, H., Takebayashi, A., Kodama, Y., Nakamura, K., Ito, H., Arino, Y., Fujita, T., Takahashi, H., Ambo, K. (1999). Effects of feed restriction and cold exposure on glucose metabolism in response to feeding and insulin in sheep. *J Anim Sci.* 77(9):2564-2573.
PMid:10492466

23. Leat, W.M., Kubasek, F.O., Buttress N. (1976). Plasma lipoproteins of lambs and sheep. *Q J Exp Physiol Cogn Med Sci.* 61 (3): 193-202.
<http://dx.doi.org/10.1113/expphysiol.1976.sp002353>
24. Henze, P., Bickhardt, K., Fuhrmann, H., Sallmann, H.P. (1998). Spontaneous pregnancy toxaemia (ketosis) in sheep and the role of insulin. *Zentralbl Veterinarmed A.* 45 (5): 255-266.
<http://dx.doi.org/10.1111/j.1439-0442.1998.tb00825.x>
PMid:9719756
25. Piccione, G., Alberghina, D., Marafioti, S., Giannetto, C., Casella, S., Assenza, A., Fazio, F. (2012). Electrophoretic serum protein fraction profile during the different physiological phases in Comisana ewes. *Reprod Domest Anim.* 47(4): 591-595.
<http://dx.doi.org/10.1111/j.1439-0531.2011.01925.x>
PMid:21988675
26. Pernthaner, A., Baumgartner, W., Jahn, J., Plautz, W., Angel, T. (1993) The hematologic parameters, concentrations of minerals and metabolic products and activities of enzymes in sheep. *Berl Munch Tierarztl Wochenschr.* 106 (3): 73-79.
PMid:8471013