



## Original Scientific Article

**BOVINE WHEY SUPPLEMENTATION IN A HIGH-FAT DIET FED RATS  
ALLEVIATED OFFSPRING'S CARDIAC INJURY**

Eman Mohammed Emara, Hassan Ibrahim El-Sayyad, Heba Atef El-Ghaweet

*Department of Zoology, Faculty of Science, Mansoura University, Mansoura, Egypt*

Received 5 February 2021; Received in revised form 6 August 2021; Accepted 18 August 2021

**ABSTRACT**

The research study determined the effect of bovine whey supplementation in rats fed on high-fat diet on occurrence of myocardium damage and disfunction in its offspring. Eighty virgin female rats (*Rattus norvegicus*) (100-110 g body weight) were used for this study. Following mating, the pregnant rats were categorized into four groups: control, whey supplemented (W), high-fat diet (FD) and high-fat diet and whey supplemented group (FD+W). Whey supplementation alone or in combination with a high-fat diet was administered every other day during the gestation and lactation period. Offspring rats at the age of 1, 7, 14 and 21-day post-partum were sacrificed and their hearts were processed for histological, p53 immunohistochemistry, transmission electron microscopy and biochemical markers for cell damage. Offspring from the FD+W group exhibited improvement of the myocardium histological picture. Moreover, there was a lower accumulation of lipid deposits and regular organization of cardiomyocyte bands and intercalated discs. A lower p53 immune reaction and lower single strand DNA damage was noticed. The levels of the antioxidant enzymes (SOD and catalase) in the myocardium were increased, whereas the contents of IL6, MDA and caspase-3 were decreased, resulting in a reduction in inflammation and cell death. In conclusion, supplementation of whey to mother rats fed with high-fat diet alleviated the markers of cardiomyocyte injury in its offspring due to its antioxidant effect.

**Key words:** high-fat diet, rats, myocardium, offspring, whey**INTRODUCTION**

Nearly 2 billion individuals worldwide are overweight, with more than half of them classed as obese, and nearly one-third of children suffered from stunted growth, and development (1). Consumption of a higher-fat-diet during pregnancy and breast-feeding period resulted in cardiac hypertrophy and increased storage of extracellular matrix as well as down-regulation of a micro-RNA subset in offspring (2). High-fat diet triggered impairment of cardiac insulin signalling resulting in insulin resistance, metabolic disturbances and mitochondrial dysfunction in both pregnancy and lactation period (3). Mice offspring whose mothers

were supplemented with a high-fat diet during pregnancy gained higher weight, had higher body length, glucose intolerance, and insulin resistance (4). Hyperlipidemic mice had higher left ventricular size which was associated with mitochondrial abnormalities and reduced oxygen consumption in the first and second generation of their neonates (5).

Bovine whey is a residual liquid of the processing of cheese, casein and yoghurt production with a pH of 5.9 - 6.6. It is composed of two main proteins,  $\beta$ -lactoglobulin ( $\beta$ -lg) and  $\alpha$ -lactalbumin ( $\alpha$ -la) which inhibit angiotensin-converting enzyme (ACE) and have antimicrobial, anticarcinogenic, and antihypercholesterolemic activity (6). Whey is also containing many bioactive fractions including glycomacropeptide,  $\beta$ -lactoglobulin,  $\alpha$ -lactalbumin, and lactoferrin, with numerous health benefits for cancer, infections, and inflammation (7). Whey supplementation alleviated cardiomyopathy and lung inflammation which was associated with increased immunohistochemical staining of caspase-3, cyclo-oxygenase-2 and induced nitric oxide synthase in a high-fat diet mother rat (8). There is no available research on the medicinal value of

*Corresponding author:* Prof. Hassan Ibrahim El-Sayyad, PhD*E-mail address:* elsayyad@mans.edu.eg*Present address:* Department of Zoology, Faculty of Science, Mansoura University, Mansoura, Egypt*Phone:* +201003179778

**Copyright:** © 2022 Emara E.M. This is an open-access article published under the terms of the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Competing Interests:** The authors have declared that no competing interests exist.

**Available Online First:** 4 March 2022**Published on:** 15 March 2022<https://doi.org/10.2478/macvetrev-2022-0017>

whey in the treatment of hyperlipidemia-related neonatal myocardial injury. The current research was undertaken to determine the alleviation role of bovine whey on the structural and functional components of offspring cardiomyocytes that had been damaged by a high-fat diet.

## MATERIAL AND METHODS

The research was approved by the Ethical Committee of the National Institute of Health and Faculty of Science, Mansoura University, Egypt, and was conducted in compliance with the regulation for laboratory animal use. The official protocol number was Sci-Z-P-2021-44.

### *Preparation of a high-fat diet*

The high-fat diet consisted of 20% lamb fat and other standard dietary components. The rats were being fed with this diet for four months before conception and during the entire time of gestation and lactation (Table 1).

### *Whey syrup supplementation*

Fresh bovine whey was daily provided from the Dairy Product Lab, Faculty of Agriculture, Mansoura University, Egypt. Each mother was administered with two doses of 2 ml by intragastric infusion, once at 8 AM and then at 2 PM, every other day. Whey syrup was administered according to the study carried out by Kandil et al. (9).

### *Experimental work*

Eighty virgin female rats (*Rattus norvegicus*) (100-110 g body weight) were obtained from the Ministry of Health, Egypt, and were acclimatized.

The zero date of gestation was noted by visualizing sperm in the vaginal smears in the next morning after mating. The pregnant rats were categorized into four groups (n=10): control, whey supplemented (W), high-fat diet (FD) and high-fat diet and whey supplemented group (FD+W). At 21 days-post-partum, the rats were fastened overnight and euthanized using chloral hydrate (300 mg/kg body weight) the following day. Maternal blood was collected at 10 AM, serum was separated by centrifugation, and then stored in refrigerator.

### *Light microscopy*

The heart ventricles of 1-, 7-, 14- and 21-day-old offspring were fixed in 10% phosphate-buffered formalin (pH 7.4) and processed for histological investigations. Serial 5 µm thick sections were cut, stained with hematoxylin and eosin, and investigated under bright- field light Olympus microscope.

### *Transmission electron microscopy*

Additional specimens were fixed in 2.5% phosphate-buffered glutaraldehyde (pH 7.4) and post-fixed in 1% phosphate-buffered osmium tetroxide. The specimens were then dehydrated in higher grades of ethyl alcohol, cleared in propylene oxide, and embedded in epoxy resin. Ultrathin sections were cut with a LKB Ultratome IV and mounted on grids. They were stained with uranyl acetate and lead citrate, and visualized on a Joel 100CX1 transmission electron microscope (Mansoura University, Egypt).

### *P53 immunohistochemistry*

Formalin-fixed and paraffin-embedded myocardial tissue sections (5 µm) were mounted on polylysine-coated glass slides and were kept at room

**Table 1.** Percentages of food components

Ingredient (%)	Control	High fat diet
Crude protein	17	17
Lamb fat	5	20
Cellulose	17	12
Casein	10	10
Vitamins	7	7
Minerals	4	4
Sucrose	10	10
Carbohydrate	10	10
Corn starch	10	10
Maltodextrin	10	-

temperature. The myocardial tissue sections of 21-day old offspring were incubated with antibodies against p53 (Thermo fisher scientific, Fremont, CA, USA). The immune stained tissue sections were counter-stained with hematoxylin, investigated, observed, and then photographed under a bright-field light microscope. Image processing was performed at 40X objective of the digital camera fixed on the Olympus microscope with 1/2 X frame adaptor. The images were analyzed using Video Test software (Russia) on an Intel Core 5 based computer and the percentage area was calculated and reported.

#### *Single cell gel electrophoresis (Comet assay)*

The myocardium of 21-day-old offspring were lysed with 1% sodium succinate, 2.5 M NaCl, 100 mM Na<sub>2</sub>EDTA, 10 mM Tris-HCl. The DNA was allowed to unwind after transferring to a buffer composed of 300 mM NaOH and 1 mM EDTA (pH 13). After 20 minutes of electrophoresis at 300 mA and 0.7 V/cm, the samples were neutralized with 0.4 M Tris-HCl buffer (pH 7). The slides were neutralized with tris buffer and stained with 100  $\mu$ L of a 2  $\mu$ g/ml ethidium bromide. The abnormal DNA strands lost their compact form and floated out of the nucleus into the agarose suspension with a low melting point. Fifty cells per each slide were analyzed using a Leitz Orthoplan (Wetzlar, Germany) fluorescence microscope with the aid of an automatic digital analysis system. Both tail length and DNA intensity were measured using image analysis software (10).

#### *Biochemical assays*

Maternal rats' serum was stored in a refrigerator. The heart tissue of 21-day-old offspring was homogenized in 10% ice-cold 2.5 mM-Tris buffer (pH 7.5) and centrifuged at 14,000 g for 15 minutes at 4 °C. The supernatant was stored in a refrigerator.

Total cholesterol (TC) (11), triglycerides (TG) (12), and high-density lipoproteins (HDL) (13) were determined in the maternal serum. The low-density lipoproteins (LDL) were calculated from the total concentrations of cholesterol (TC), HDL-cholesterol, and triglycerides according to Friedewald et al. (14). The serum levels of leptin (CSB-E07433r), insulin (CSB-E05070r), adiponectin (CSB-E07271r) was determined by using ELISA kit of Cusabio Biotech Company (Huston, USA). Troponin I is determined by ELISA Kit (ab246529), while alkaline phosphatase (ALP) (ab83369) and lactic dehydrogenase (LDH)

(ab197000) was determined colorimetrically according to the instruction of Abcam Biotech Company (Cambridge, United Kingdom).

The heart samples of 21-day-old offspring were weighed, then homogenized in 10% ice-cold 2.5 mM-Tris buffer (pH 7.5) and centrifuged at 14,000 g for 15 minutes at 4 °C. The supernatants were stored in a refrigerator.

Superoxide dismutase was determined by a reduction of nitro blue tetrazolium (NBT) with superoxide radicals to a blue formazan color assayed colorimetrically at 560 nm (15). Catalase was assayed by the degradation of hydrogen peroxide and the reaction product of non-reacted hydrogen peroxide was measured at 550 nm (16). Malondialdehyde (MDA) was determined by measuring the thiobarbituric acid concentration (17). Interleukin 6 was estimated by using Enzyme-linked Immunosorbent Assay Kit of Cloud-Clone Corp, (catalogue No. SEA079Ra). Also, caspase-3, was determined colorimetrically by using a Stressgen kit (catalog No. 907-013) based on the hydrolysis of the peptide substrate acetyl-Asp-Glu-ValAsp p-nitroanilide (Ac-DEVD-pNA) and the resulting formation of the p-nitroaniline (PNA) was measured spectrophotometrically at 405 nm and compared with a standard curve.

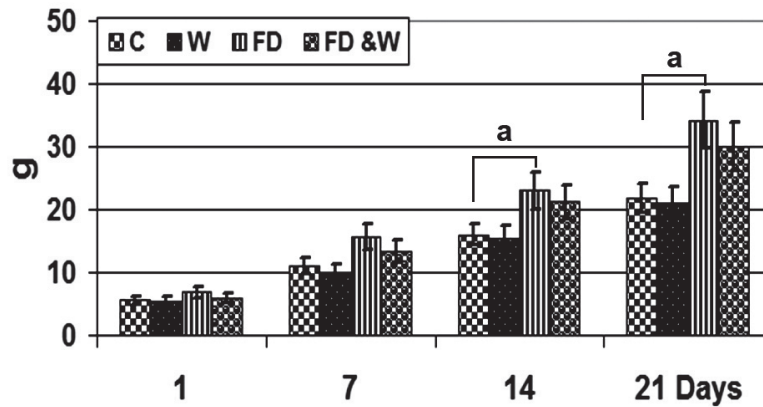
#### *Statistical analysis*

The statistical analysis was performed by SPSS (version 13). Two-way ANOVA and post hoc analysis were employed between the control and experimental groups as well as between the high-fat diet and high-fat diet plus whey group. Data are presented as means  $\pm$  standard error (SE). The t-test was calculated and considered statistically significant at  $p < 0.05$ .

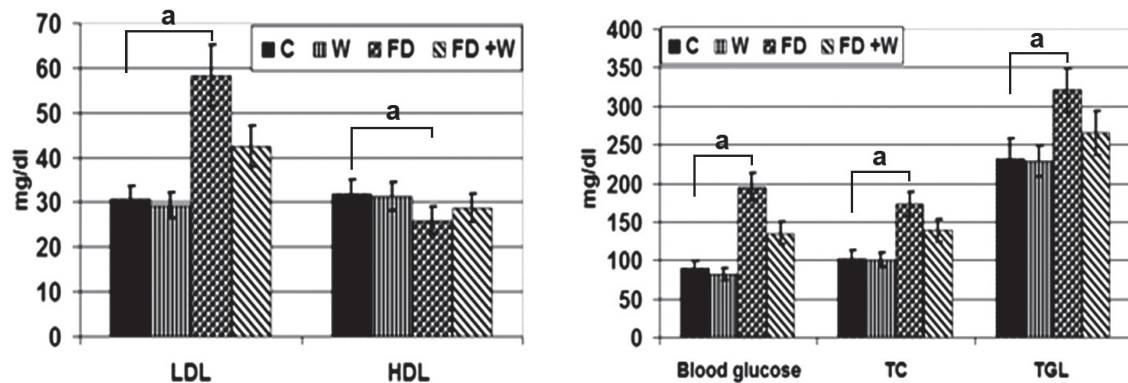
## RESULTS

#### *Body weight gain*

The 1-, 7-, 14- and 21-day-old offspring from the FD group gained significantly higher body weight ( $6.00 \pm 0.75$ ,  $15.60 \pm 1.51$ ,  $23.00 \pm 2.95$  and  $34.20 \pm 4.49$ , respectively) compared to the control ( $5.58 \pm 0.47$ ,  $11.00 \pm 1.09$ ,  $15.50 \pm 1.35$  and  $21.80 \pm 1.73$ , respectively). The 1-, 7-, 14 and 21-day-old offspring from the FD+W had insignificant increase in body weight compared to the control ( $5.80 \pm 0.54$ ,  $13.40 \pm 1.40$ ,  $21.20 \pm 2.93$  and  $30.00 \pm 2.71$ , respectively) (Fig. 1).



**Figure 1.** Body weight gain of offspring from mother rats fed with a high-fat diet with or without whey supplementation. Each value is the mean  $\pm$  SE (n=5/group). Abbreviations: C - control group; W - whey supplemented group; FD - high-fat diet group; FD+W - high-fat diet and whey supplementation group. The symbol 'a' marks significant difference of FD compared to control (P<0.05)



**Figure 2.** Serum LDL, HDL, blood glucose, TC and TGL level of mother rats fed with a high-fat diet with or without whey supplementation. Each value is the mean  $\pm$  SE (n=5/group). Abbreviations: C - control group; W - whey supplemented group; FD - high-fat diet group; FD+W - high-fat diet and whey supplementation group; LDL - low-density lipoproteins; HDL - high-density lipoproteins; TC - total cholesterol; TGL - triglycerides. The index "a" marks significant difference of FD compared to control at P<0.05

**Table 2.** Serum biochemical markers of mother rats fed with a high-fat diet with or without whey supplementation

	Insulin ( $\mu$ U/ml)	Leptin (pg/ml)	Adiponectin (ng/ml)	Troponin I (pg/ml)	ALP (U/L)	LDH (mU/ml)
C	311.90 $\pm$ 13.36	266.70 $\pm$ 29.22	25.40 $\pm$ 1.60	63.30 $\pm$ 3.45	212.90 $\pm$ 7.96	158.90 $\pm$ 10.42
W	321.10 $\pm$ 12.89	343.40 $\pm$ 29.17	26.04 $\pm$ 1.59	56.70 $\pm$ 3.62	207.60 $\pm$ 7.94	160.90 $\pm$ 9.50
FD	466.30 $\pm$ 13.11 <sup>a</sup>	1605.30 $\pm$ 35.61 <sup>a</sup>	14.80 $\pm$ 1.59 <sup>a</sup>	96.20 $\pm$ 3.34 <sup>a</sup>	379.90 $\pm$ 8.08 <sup>a</sup>	431.10 $\pm$ 9.63 <sup>a</sup>
FD+W	428.70 $\pm$ 12.84 <sup>a</sup>	1248.70 $\pm$ 29.33 <sup>a,b</sup>	18.10 $\pm$ 1.59 <sup>a</sup>	87.60 $\pm$ 3.36 <sup>a</sup>	359.90 $\pm$ 8.07 <sup>a</sup>	395.10 $\pm$ 9.68 <sup>a</sup>
F-test (FD differences from C)	35.018	360.672	12.018	30.210	133.551	229.169
F-test (FD differences from FD+W)	4.22	59.74	2.17	3.33	3.07	9.87
P<0.05	S.	S.	S.	S.	S.	S.

Each value is the mean  $\pm$  SE (n=5/group). Abbreviations: C - control group; W - whey supplemented group; FD - high fat diet group; FD+W - high fat diet and whey supplementation group; ALP - alkaline phosphatase; LDH - lactic dehydrogenase. The symbol "a" in superscript marks significant difference between FD and control. Symbol "b" in superscript marks significant difference between FD and FD+W (P<0.05)



*Maternal serum biochemical changes*

The W group had non-significant lower serum levels of LDL, triglycerides, cholesterol and glucose (Fig. 2). Whey administration has also reduced the rapid increase of insulin, leptin, troponin I, alkaline phosphatase and lactic dehydrogenase. Furthermore, mother rats of the FD+W showed decreased serum level of adiponectin (Table 2).

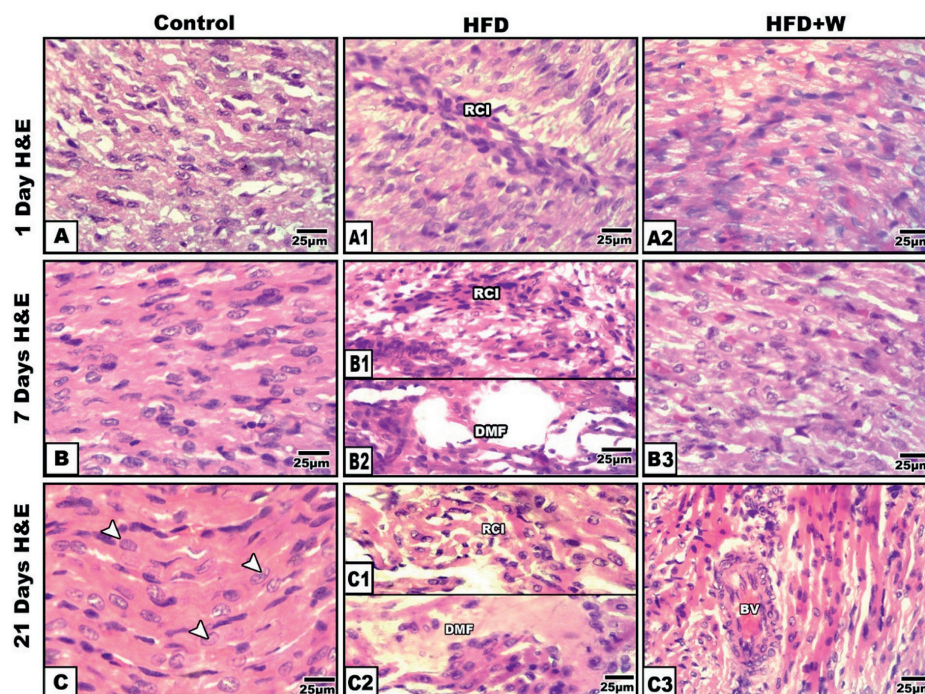
*Biochemical observations in myocardium of 21-day-old offspring*

In the FD group, the neonate myocardial tissues showed significantly lower SOD and catalase ( $12.19 \pm 0.96$  and  $21.41 \pm 1.31$ , respectively) compared to the control ( $17.22 \pm 0.96$  and  $30.59 \pm 0.97$ , respectively). The MDA content was significantly higher ( $5.95 \pm 0.49$ ) than the control ( $3.33 \pm 0.46$ ).

**Table 3.** Biochemical markers of myocardium of 21-day-old offspring with or without whey supplementation

	SOD (U/mg p)	Catalase (mU/mg p)	MDA (nmol/mg p)	IL6 (pg/mg p)	Caspase-3 (ng/mg p)
C	$17.22 \pm 0.96$	$30.59 \pm 0.97$	$3.33 \pm 0.46$	$27.18 \pm 1.64$	$3.20 \pm 0.26$
W	$16.75 \pm 0.97$	$30.11 \pm 1.28$	$3.27 \pm 0.33$	$27.37 \pm 1.61$	$3.01 \pm 0.23$
FD	$12.19 \pm 0.96^a$	$21.41 \pm 1.31^a$	$5.95 \pm 0.49^a$	$70.36 \pm 1.74^a$	$5.70 \pm 0.50^a$
FD+W	$13.70 \pm 0.49^a$	$24.03 \pm 0.84^a$	$4.90 \pm 0.18^a$	$45.78 \pm 1.63^{a,b}$	$4.50 \pm 0.20^a$
F-test (FD differences from C)	8.461	16.440	11.426	151.572	15.408
F-test (FD differences from FD+W)	0.83	2.83	4.01	105.93	5.01
P<0.05	S.	S.	S.	S.	S.

Each value is the mean  $\pm$  SE (n=5/group). Abbreviations: C - control group; W - whey supplemented group; FD - high-fat diet group; FD+W - high-fat diet and whey supplementation group; IL6 - interleukin 6; MDA - malondialdehyde; SOD - superoxide dismutase. The symbol "a" in superscript signifies significant difference between FD and control (P<0.05); The symbol "b" in superscript signifies significant difference between FD and FD+W (P<0.05)



**Figure 3.** Photomicrographs of histological sections of heart ventricle of rat offspring. 1 day-old offspring (A-A2); Control (A-C); 1- & 21-day-old offspring of mother fed with a high fat diet showing fragility of muscle fibers, hyaline necrosis and leukocytic infiltration (A1-B1&B2-C1&C2); 1-, 7- & 21-day old offspring of high fat-diet mother supplemented with whey showing improved orientation of muscle fibers (A2, B3 & C3). H&E. Abbreviations: DMF - degenerated muscle fibers; RCI - round cells infiltration; BV - blood vessel; RCI - round cells infiltration; IH - internal hemorrhage

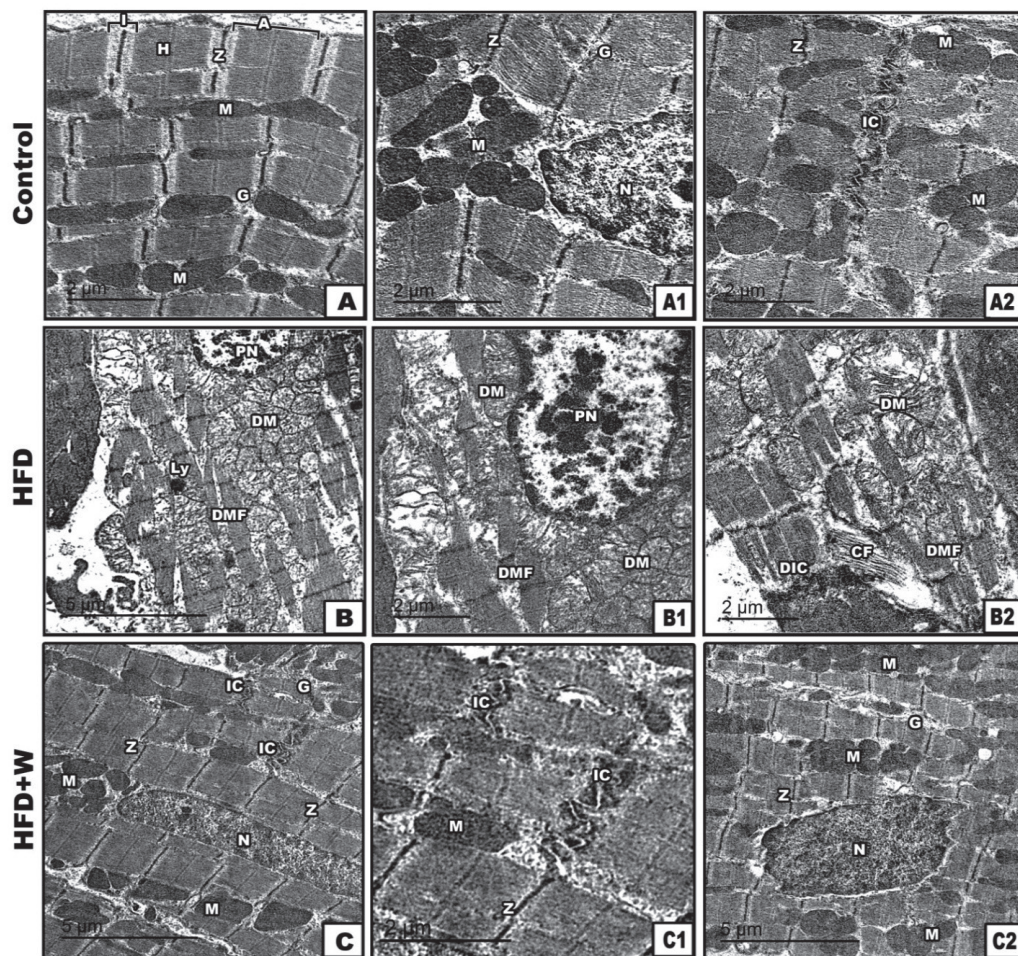
In addition, the myocardial contents of IL6 and caspase-3 were significantly higher ( $70.36 \pm 1.74$  and  $5.70 \pm 0.50$ , respectively) than the control ( $27.18 \pm 1.64$  and  $3.20 \pm 0.26$ , respectively).

In the FD+W, there was a significantly lower MDA content ( $4.39 \pm 0.18$ ) than the control ( $3.33 \pm 0.46$ ). The level of IL6 was significantly lower ( $45.78 \pm 1.63$ ) compared to the control ( $27.18 \pm 1.64$ ). In addition, the caspase content had non-significantly higher ( $4.50 \pm 0.20$ ) value compared to the control ( $3.20 \pm 0.26$ ). The myocardial level of IL6 was significantly lower in FD+W ( $45.00 \pm 1.63$ ) compared to the FD group ( $70.36 \pm 1.74$ ). The other assayed parameters were non-significantly changed (Table 3).

#### Light and ultrastructural observation

The 1-, 7- and 21-day-old offspring showed disorganization of the myocardial fibers compared to the control (Fig. 3, A-C). Focal leukocyte infiltration was observed between the muscle fibers. Numerous foci of necrotic fibers have been associated with hyalinization, fragmentation, sarcoplasmic vacuolation, and pyknosis (Fig. 3 A1, B1, B2, C1 and C2). Offspring from HFD and W group had improved histopathological findings on the myocardial muscle (Fig. 3 A2, B3 and C3).

The 21- day-old offspring of FD mothers had irregular corrugated sarcolemma with pyknotic nuclei characterized by tightly compacted heterochromatin.



**Figure 4.** Transmission electron micrographs of myocardium of 21-day-old rat. Control showing regular arranged muscle fibers with characteristic A-&I-band, Z-line & H-zone. Abundant mitochondria appears distributed in between muscle fibers (A-A2); 21-day-old rat from mothers fed with high fat-diet showing degenerated muscle fibers, damaged mitochondria, pyknotic myocyte nuclei (B) and degenerated intercalated disc (B-B2); 21 day- old offspring from mothers fed with high fat diet supplemented with whey showing ameliorated muscle fibers with regularly oriented muscle bands, cardiomyocyte nuclei and abundant mitochondria in between muscle fibers (C-C2). Abbreviations: CF - collagen fibers; DM - degenerated mitochondria; DMF - degenerated muscle fibers; DIC - degenerated intercalated disc; G – granules; H - H band; IC - intercalated disc; LY - lysosome; M - mitochondria; N - nucleus; PN - pyknotic nuclei; Z - Z-line



The intercalated disks were abnormally folded. The interfibrillar mitochondria were atrophied and aggregated rather than being radially distributed among the fibrils (Fig. 4 B-B2). Offspring of FD+W rats had improved muscle fiber. The muscle fibers had normal structure. The muscle bands were regularly oriented and both mitochondria and myocyte nuclear chromatin structures were more consistent (Fig. 4 C-C2) compared to the regular and normal characteristic pattern of the control (Fig. 4 A-A2).

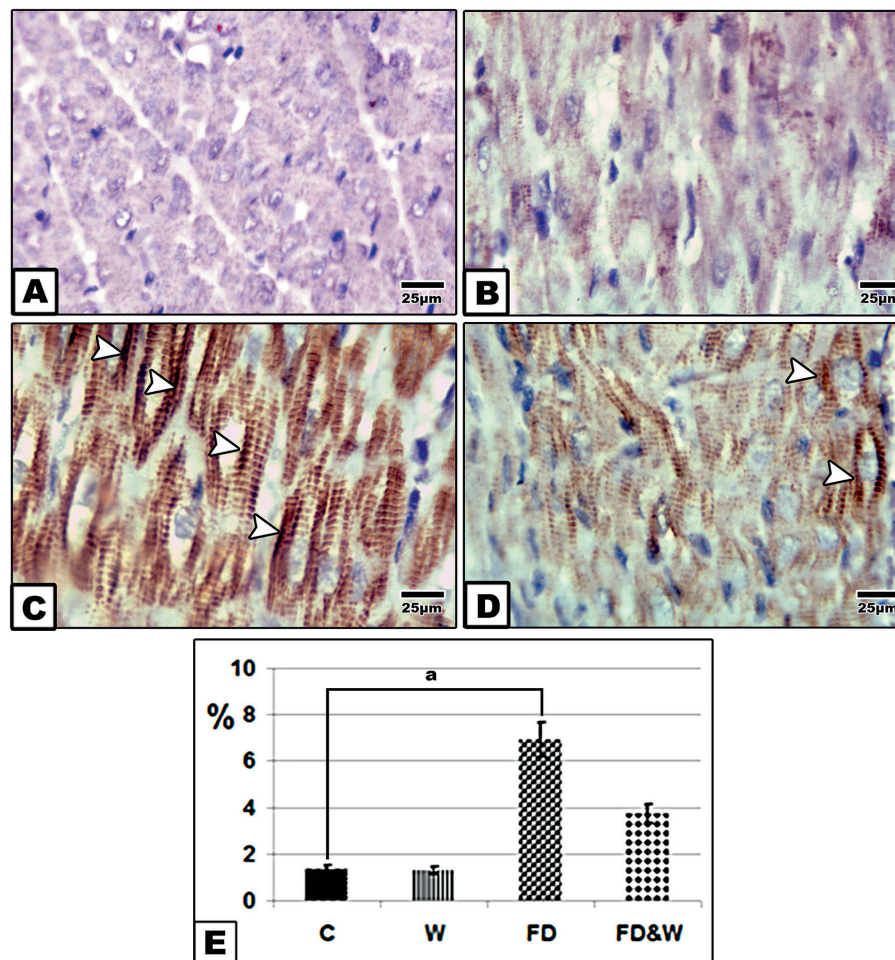
#### Immunohistochemistry of P53

The myocardium of 21-day-old offspring from the FD group revealed overexpression of P53

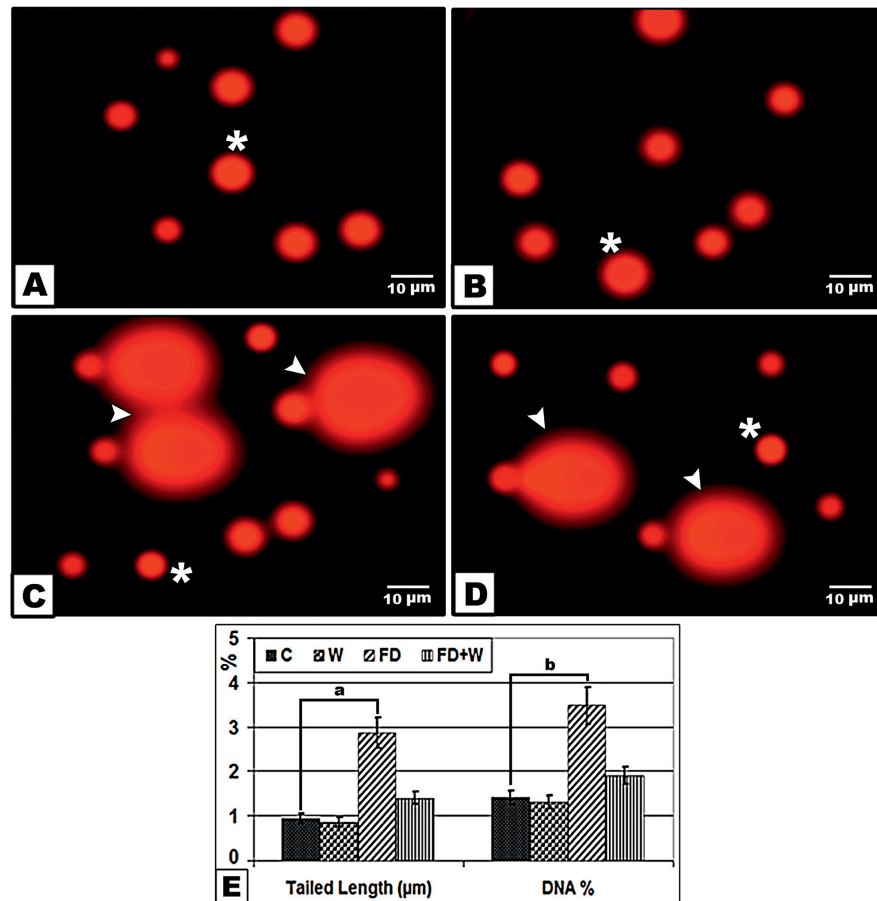
compared to the offspring from the other groups (Fig. 5 A-D). Hence, the FD group offspring had significantly higher immune reaction compared to the other groups (Fig. 5 E).

#### Single DNA fragmentation (Comet assay)

Compared to the control, single strand DNA showed increased detached cardiomyocyte tail and DNA concentration reflecting DNA damage in 21-day-old offspring of mother ingested high-fat diet. However, those maternally supplemented whey plus fat diet showed marked decrease of DNA damage within the cardiomyocytes (Fig. 6).



**Figure 5.** Photomicrographs of formalin fixed histological sections of heart immunohistochemical stained with P53 of 21-day-old rat. Control and whey supplemented group showing decreased immunohistochemical reaction of P53 (A&B); Offspring of mother fed with a high-fat-diet group showing increase dark brown immunohistochemical reaction of P53 in muscle fibers manifesting cell death (C); Offspring of mother fed with a high fat-diet supplemented with whey showing decreased immunohistochemical reaction of P53 (D). Arrow head indicating reaction of P53. Histogram illustrating the percentages of immunohistochemical reaction expression of P53 in myocardium of offspring of mother fed with high fat-diet and improved in the group fed with a high fat diet and supplemented whey plus the fat diet (E). The symbol “a” marks difference compared to the control at  $P < 0.05$



**Figure 6.** Comet assay of heart ventricle of 21-day-old rat of mother from mothers fed with a high fat-diet with or without whey supplementation. Control (A); Whey supplementation (B); High fat-diet (C); High fat-diet supplemented with whey (D). The symbol (\*) marks normal cells. Arrow head points detached tail region (DNA damage). Chart illustrating tailed length and DNA % of damaged cardiomyocytes (E). The symbols “a” and “b” mark significant differences between the high fat-diet and control ( $P < 0.05$ )

## DISCUSSION

The current study revealed that offspring of FD mothers gained more body weight and showed symptoms of obesity compared to the offspring from mothers fed with normal diet. This trend was first noticed at parturition and continued throughout the suckling period. These findings are conflicting with the report of Ribaroff et al. (18) stating that maternal high-fat diet resulted in increased body weight only at weaning. Butruille et al. (19) reported that the increased body weight of rat offspring from obese mothers was a result of increased visceral adiposity.

In addition, histopathological abnormalities were detected on the myocardium of the FD offspring. The changes were characterized by hyalinized necrosis of the cardiomyocytes

associated with the dense leukocyte cell infiltration and disorganized muscle fibers. At ultrastructural level, the distorted sarcolemma was characterized with almost missing Z and I bands, deformed intercalated discs and the presence of pyknotic nuclei. The interfibrillar mitochondria were atrophied and clumped instead of being radially distributed between the fibrils. Interfibrillar lipid deposits have been clearly identified. The current results are consistent with the findings of Mdaki et al. (3) who reported impaired cardiac structure and function in rat offspring from mothers fed with high-fat diet. Guzzardi et al. (20) has reported a thicker left ventricular posterior wall and disrupted diastolic contraction in pig neonate from obese mothers.

Mother rats fed with a high-fat diet had higher serum levels of LDL, triglycerides, total cholesterol



and blood glucose that may predict early signs of diabetes. This was highly indicative of myocardial damage in offspring and could be predictive to the onset of diabetes. The increased oxidation of fatty acids and release of free radicals (21) leading to maternal myocardial fibrosis (8) could be attributed to the significantly higher serum levels of troponinI, LDH and ALP which was highly predictive of cardiotoxicity in the mothers. This is compliant with the report of Magalhães et al. (22) on diabetic rats fed with high-fat diet.

The cardiomyocyte damage of offspring from FD mothers was characterized by significantly lower antioxidant enzymes (SOD and catalase) resulting in oxidative stress (high MDA levels). It was found that this led to increase the overproduction of the superoxide ( $O_2^-$ ) and oxygen species from the damaged interfibrillar mitochondria. The same was also reported for myocardial tissues of rats fed with a high-fat diet (23).

The high expression of P53 in cardiomyocytes of FD offspring was also reported by Attia and Taha (24) in apoptotic cells of diabetic rats. These findings were confirmed by the comet assay which detected increased myocardial caspase-3. Rat offspring from mothers fed with a high cholesterol and triglyceride diet were more likely to develop type 2 diabetes (25). It is also known that troponin-I is associated with managing of the actin thin filament within muscle cells. Its function is carried out in conjugation with the alkaline phosphatase. The increased activities of both enzymes indicated cardiomyocyte damage (26). You et al. (27) reported that the elevation of alkaline phosphatase predicted cardiovascular disease.

The FD+W group offspring had regular cardiomyocyte structure which is coinciding with the lower biochemical markers for cardiomyocyte damage. Increased levels of the myocardial SOD and catalase coincided with the decreased oxidative stress. Also, whey was found to decrease the IL6 which facilitated myocardial inflammation, and caspase-3 and troponin-I involved in the cardiomyocyte cell death. Whey supplementation was found to improve the histological structure and function of aging cardiac tissue (28) by regulating the renin-angiotensin-aldosterone system (29), insulin levels in mice (30) and by increasing cardiac levels of the antioxidant enzyme activities of GPx and GSH in cardiomyocytes (31). In addition, high-fat diet was found to decrease the gut microbiota in mother rats (32). Administration of probiotic alleviated the reduction of triglycerides, cholesterol and LDL

of eighteen-month-old *Macaca fuscata* offspring from mothers fed with a high-fat diet (33).

## CONCLUSION

Whey exhibited antioxidant activity and contained adequate amounts of protein and lactobacilli. Supplementation of whey with a high-fat diet facilitated upregulation of serum levels of adiponectin and alleviated the markers of cardiomyocyte injury. Whey supplementation improved the antioxidant defence and reduced the oxidative stress in the myocardium of rat offspring from mothers fed with high-fat diet mother.

## CONFLICT OF INTEREST

The authors declared that they have no potential conflict of interest with respect to the authorship and/or publication of this article.

## ACKNOWLEDGEMENTS

This research was supported by the Faculty of Science, Mansoura University, Egypt.

## AUTHORS' CONTRIBUTIONS

HIEI-S designed the experiment, formatted the data, revised, and constructed the manuscript. HAEI-G constructed the first manuscript version, performed the histopathology investigations, and statistical analysis. EME have performed the experimental work and prepared the histopathology and transmission electron microscopy specimens.

## REFERENCES

- Hoffman, D.J., Powell, T.L., Barrett, E.S., Hardy, D.B. (2021). Developmental origins of metabolic diseases. *Physiol Rev.* 101(3): 739-795.  
<https://doi.org/10.1152/physrev.00002.2020>  
PMid:33270534
- Siddeek, B., Mauduit, C., Chehade, H., Blin, G., Liand, M., Chindamo, M. et al. (2019). Long-term impact of maternal high-fat diet on offspring cardiac health: role of micro-RNA biogenesis. *Cell Death Discov.* 5, 71.  
<https://doi.org/10.1038/s41420-019-0153-y>  
PMid:30854230 PMCID:PMC6397280

3. Mdaki, K.S., Larsen, T.D., Wachal, A.L., Schimelpfenig, M.D., Weaver, L.J., Dooyema, S.D. et al. (2016). Maternal high-fat diet impairs cardiac function in offspring of diabetic pregnancy through metabolic stress and mitochondrial dysfunction. *Am J Physiol Heart Circ Physiol.* 310, H681-H692.  
<https://doi.org/10.1152/ajpheart.00795.2015>  
PMid:26801311 PMCID:PMC4867345
4. Dunn, G.A., Bale, T.L. (2009). Maternal high-fat diet promotes body length increases and insulin insensitivity in second-generation mice. *Endocrinology* 150(11): 4999-5009.  
<https://doi.org/10.1210/en.2009-0500>  
PMid:19819967 PMCID:PMC2775990
5. Ferey, J.L.A., Boudoures, A.L., Reid, M., Drury, A., Scheaffer, S., Modi, Z. et al. (2019). A maternal high-fat, high-sucrose diet induces transgenerational cardiac mitochondrial dysfunction independently of maternal mitochondrial inheritance. *Am J Physiol Heart Circ Physiol.* 316(5): H1202-H1210.  
<https://doi.org/10.1152/ajpheart.00013.2019>  
PMid:30901280 PMCID:PMC6580388
6. Chatterton, D.E.W., Smithers, G., Roupas, P., Brodtkorb, A. (2006). Bioactivity of  $\beta$ -lactoglobulin and  $\alpha$ -lactalbumin-Technological implications for processing. *Int Dairy J.* 16(11): 1229-1240.  
<https://doi.org/10.1016/j.idairyj.2006.06.001>
7. Krissansen, G.W. (2007). Emerging health properties of whey proteins and their clinical implications. *J Am Coll Nutr.* 26(6): 713S-723S.  
<https://doi.org/10.1080/07315724.2007.10719652>  
PMid:18187438
8. El-Sayyad, H.I., El-Ghawet, H.A., El-Bayomi, K.S., Emara, E. (2020). Bovine whey improved the myocardial and lung damage of mother rats fed on a high fat diet. *Stud Stem Cells Res Ther.* 6(1): 001-008.  
<https://doi.org/10.17352/sscr.000014>
9. Kandil, N.T.A.H. Sabry, D.A.M., Ashry, N.E.E., El-Sayyad, H.I.H. (2020). Therapeutic potential of whey against aging related cytological damage of adenohipophysis of rat. *East African Scholars J Agri Life Sci.* 3(9): 304-310.  
<https://doi.org/10.36349/EASJALS.2020.v03i09.002>
10. Sasaki, Y.F., Nishidate, E., Izumiyama, F., Matsusaka, N., Tsuda, S. (1997). Simple detection of chemical mutagens by the alkaline single-cell gel electrophoresis (Comet) assay in multiple mouse organs. *Mutat Res.* 391(3): 215-231.  
[https://doi.org/10.1016/S1383-5718\(97\)00073-9](https://doi.org/10.1016/S1383-5718(97)00073-9)
11. Deeg, R., Ziegenhorn, J. (1983). Kinetic enzymic method for automated determination of total cholesterol in serum. *Clin Chem.* 29(10): 1798-1802.  
<https://doi.org/10.1093/clinchem/29.10.1798>  
PMid:6577981
12. Fossati, P., Prencipe, L. (1982). Serum triglycerides determined colorimetrically with an enzyme that produces hydrogen peroxide. *Clin Chem.* 28(10): 2077-2080.  
<https://doi.org/10.1093/clinchem/28.10.2077>  
PMid:6812986
13. Grove, T.H. (1979). Effect of reagent PH on determination of the high-density lipoprotein cholesterol by precipitation with sodium phosphotungstate-magnesium. *Clin Chem.* 25(4): 560-564.  
<https://doi.org/10.1093/clinchem/25.4.560>  
PMid:38018
14. Friedewald, W.T., Levy, R.I., Fredrickson, D.S. (1972). Estimation of low density lipoprotein cholesterol in plasma without use preparative ultracentrifuge. *Clin Chem.* 18(6): 499-502.  
<https://doi.org/10.1093/clinchem/18.6.499>  
PMid:4337382
15. Niskikimi, M., Rao, N., Yaki, K. (1972). The occurrence of superoxide anion in the reaction of reduced phenazinemethosulfate and molecular oxygen. *Biochem Biophys Res Commun.* 46(2): 849-854.  
[https://doi.org/10.1016/S0006-291X\(72\)80218-3](https://doi.org/10.1016/S0006-291X(72)80218-3)
16. Bock, P.P., Kramer, R., Pavelka, M. (1980). Peroxisomes and related particles. In M. Alfert, W. Beermann, L. Goldstein, K.R. Porter, P. Sitte (Eds.), *Cell Biology Monographs* 7 (pp. 44-74). Springer, Berlin  
[https://doi.org/10.1007/978-3-7091-2055-2\\_2](https://doi.org/10.1007/978-3-7091-2055-2_2)
17. Ohkawa, H., Ohishi, N., Yagi, K. (1979). Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal Biochem.* 95(2): 351-358.  
[https://doi.org/10.1016/0003-2697\(79\)90738-3](https://doi.org/10.1016/0003-2697(79)90738-3)
18. Ribaroff, G.A., Wastnedge, E., Drake, A.J., Sharpe, R.M., Chambers, T.J.G. (2017). Animal models of maternal high fat diet exposure and effects on metabolism in offspring: a meta-regression analysis. *Obes Rev.* 18(6): 673-686.  
<https://doi.org/10.1111/obr.12524>  
PMid:28371083 PMCID:PMC5434919
19. Butruille, L., Marouze, L., Pourpe, C., Oger, F., Lecoutre, S., Catheline, D. et al. (2019). Maternal high-fat diet during suckling programs visceral adiposity and epigenetic regulation of adipose tissue stearoyl-CoA desaturase-1 in offspring. *Int J Obes (Lond).* 43(12): 2381-2393.  
<https://doi.org/10.1038/s41366-018-0310-z>  
PMid:30622312
20. Guzzardi, M.A., Liistro, T., Gargani, L., Ait Ali, L., D'Angelo, G., Rocchiccioli, S. et al. (2018). Maternal obesity and cardiac development in the offspring: Study in human neonates and minipigs. *JACC Cardiovasc Imaging.* 11(12): 1750-1755.  
<https://doi.org/10.1016/j.jcmg.2017.08.024>  
PMid:29153568

21. Giacco, F., Brownlee, M. (2010). Oxidative stress and diabetic complications. *Circ Res.* 107(9): 1058-1070. <https://doi.org/10.1161/CIRCRESAHA.110.223545> PMID:21030723 PMCID:PMC2996922
22. Magalhães, D.A., Kume, W.T., Correia, F.S., Queiroz, T.S., Allebrandt Neto, E.W., Santos, M.P.D. et al. (2019). High-fat diet and streptozotocin in the induction of type 2 diabetes mellitus: a new proposal. *An Acad Bras Cienc.* 91(1): e20180314. <https://doi.org/10.1590/0001-3765201920180314> PMID:30916157
23. Xiang, L., Zhang, Q., Chi, C., Wu, G., Lin, Z., Li, J. et al. (2020). Curcumin analog A13 alleviates oxidative stress by activating Nrf2/ARE pathway and ameliorates fibrosis in the myocardium of high-fat-diet and streptozotocin-induced diabetic rats. *Diabetol Metab Syndr.* 12, 1. <https://doi.org/10.1186/s13098-019-0485-z> PMID:31921358 PMCID:PMC6947902
24. Attia, H.M., Taha, M. (2018). Protective effect of captopril on cardiac fibrosis in diabetic albino rats: a histological and immunohistochemical study. *Benha Med J.* 35(3): 378-385. [https://doi.org/10.4103/bmfj.bmfj\\_122\\_18](https://doi.org/10.4103/bmfj.bmfj_122_18)
25. Sheen, J.M., Yu, H.R., Tain, Y.L., Tsai, W.L., Tiao, M.M., Lin, I.C., Tsai, C.C., Lin, Y.L., Huang, L.T. (2018). Combined maternal and postnatal high-fat diet leads to metabolic syndrome and is effectively reversed by resveratrol: a multiple-organ study. *Sci Rep.* 8(1): 5607. <https://doi.org/10.1038/s41598-018-24010-0> PMID:29618822 PMCID:PMC5884801
26. Dasgupta, A., Chow, L., Wells, A., Datta, P. (2001). Effect of elevated concentration of alkaline phosphatase on cardiac troponin I assays. *J Clin Lab Anal.* 15(4): 175-177. <https://doi.org/10.1002/jcla.1023> PMID:11436198 PMCID:PMC6807912
27. You, A.H., Han, D.W., Ham, S.Y., Lim, W., Song, Y. (2019). Serum alkaline phosphatase as predictor of cardiac and cerebrovascular complications after lumbar spinal fusion surgery in elderly: A retrospective study. *J Clin Med.* 8(8): 1111. <https://doi.org/10.3390/jcm8081111> PMID:31357535 PMCID:PMC6723677
28. Al-Gebaly, A.S. (2019). Ameliorating role of whey syrup against ageing- related damage of myocardial muscle of Wistar Albino rats. *Saudi J Biol Sci.* 26(5): 950-956. <https://doi.org/10.1016/j.sjbs.2018.03.014> PMID:31303824 PMCID:PMC6600591
29. Martin, M., Kopalani, I., Jannasch, A., Mund, C., Todorov, V., Henle, T. et al. (2015). Antihypertensive and cardioprotective effects of the dipeptide isoleucine-tryptophan and whey protein hydrolysate. *Acta Physiol (Oxf).* 215(4): 167-176. <https://doi.org/10.1111/apha.12578> PMID:26297928
30. El-Shinnawy, N.A., Abd Elhalem, S.S., Haggag, N.Z., Badr, G. (2018). Ameliorative role of camel whey protein and rosuvastatin on induced dyslipidemia in mice. *Food Funct.* 9(2): 1038-1047. <https://doi.org/10.1039/C7FO01871A> PMID:29349446
31. Bartfay, W.J., Davis, M.T., Medves, J.M., Lugowski, S. (2003). Milk whey protein decreases oxygen free radical production in a murine model of chronic iron-overload cardiomyopathy. *Can J Cardiol.* 19(10): 1163-1168.
32. Mann, P.E., Huynh, K., Widmer, G. (2018). Maternal high fat diet and its consequence on the gut microbiome: A rat model. *Gut Microbes.* 9(2): 143-154. <https://doi.org/10.1080/19490976.2017.1395122> PMID:29135334 PMCID:PMC5989793
33. Pace, R.M., Prince, A.L., Ma, J., Belfort, B.D.W., Harvey, A.S., Hu, M. et al. (2018). Modulations in the offspring gut microbiome are refractory to postnatal synbiotic supplementation among juvenile primates. *BMC Microbiol.* 18, 28. <https://doi.org/10.1186/s12866-018-1169-9> PMID:29621980 PMCID:PMC5887201



