

*Original Scientific Article***MOUSE NUCLEOLUS-LIKE BODY SUPPORTS TRANSCRIPTION AND ENHANCES PROCESSING OF rRNA IN PORCINE ENUCLEOLATED OOCYTES**

Martin Morovic¹, Frantisek Strejcek¹, Michal Benc¹, Alexandra Rosenbaum Bartkova^{1,2}, Sara Stachovicova^{1,3}, Florina Popovska-Percinic⁴, Monika Dovenska⁴, Jozef Laurincik¹

¹*Faculty of Natural Sciences, Constantine the Philosopher University in Nitra,
Nabrezie Mladeze 91, 94974 Nitra, Slovak Republic*

²*Institute of Animal Physiology and Genetics, Czech Academy of Sciences,
Rumburska 89, 277 21 Libechov, Czech Republic*

³*Université Paris-Saclay, Université de Versailles Saint-Quentin-en-Yvelines, INRAE,
BREED, Jouy-en-Josas, France*

⁴*Faculty of Veterinary Medicine-Skopje, Ss. Cyril and Methodius University in Skopje,
1000 Skopje, North Macedonia*

Received 5 June 2024; Received in revised form 28 August 2024; Accepted 4 September 2024

ABSTRACT

Fully-grown mammalian oocytes contain morphologically evident but transcriptionally inactive nucleoli called “nucleolus-like bodies” (NLBs). These nuclear structures are essential for early embryonic development. Removing oocyte NLBs (enucleolation) before their activation leads to developmental failure, mainly at the time point of embryonic genome activation. The present study examined the developmental and expression dynamics of embryos derived from intra- (pig) and interspecies (mouse) nucleolus (NLB) transferred porcine oocytes after parthenogenetic activation. Activation of rDNA transcription and pre-rRNA processing in NLB re-injected embryos was observed by real-time qRT-PCR analysis, targeting the expression rates of 45S rRNA and levels of 18S rRNA. Re-injection of NLBs from mouse or porcine GV oocytes into the enucleolated MII stage porcine oocytes supported the embryonic development up to the blastocyst stage after their activation. Intra- and intergeneric NLB transferred embryos demonstrated rDNA transcription initiation at the 8-cell stage, corresponding with the embryonic genome activation in porcine parthenogenetic embryos (control). The measured levels of 18S rRNA in these experimental embryos showed delayed initiation of pre-rRNA processing at the blastocysts stage compared with the control (8-cell stage). However, the porcine embryos with re-injected mouse NLB displayed significantly higher levels of 18S rRNA than the control and experimental intraspecies group. In conclusion, NLBs from different mammal species (mouse) can enhance the quality of enucleolated porcine oocytes and thus support their early embryonic development after activation.

Key words: nucleolus-like body, oocyte, pig, mouse, nucleolotransfer

INTRODUCTION

The major genome activation (MGA) in mammalian embryos is unambiguously characterized by consecutive activation of RNA-polymerase-I, -II,

and -III transcription, resulting in successful embryonic development. Processes of rDNA transcription, mediated by RNA-polymerase-I, and the subsequent processing of rRNA to the formation of pre-ribosomal subunits, are localized in the nucleolus. The activation of rDNA transcription is also reflected at the ultrastructural level by the transformation of the dense nucleolar structure (nucleolus precursor body - NPB) into a functional tripartite nucleolus consisting of fibrillar centers, a dense fibrillar component, and the granular component (1). In contrast to MGA, rRNA synthesis increases during oocyte growth and peaks at the beginning of follicular antrum formation in early mammalian embryos. The full meiotic

Corresponding author: Prof. Frantisek Strejcek, PhD

E-mail address: fstrejcek@ukf.sk

Present address: Faculty of Natural Sciences, Constantine the Philosopher University in Nitra, Nabrezie Mladeze 91, 94974 Nitra, Slovak Republic

Phone: +421 37 6408 584

Copyright: © 2025 Morovic M. This is an open-access article published under the terms of the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Competing Interests: The authors have declared that no competing interests exist.

Available Online First: 10 January 2025

Published on: 15 March 2025

<https://doi.org/10.2478/macvetrev-2025-0011>

competence coincides with a markedly decreased transcriptional activity and formation of temporary compact fibrillar nucleolar sphere (nucleolus-like body - NLB) in nearly grown oocytes at the germinal vesicle (GV) stage (2). Immediately before germinal vesicle breakdown (GVBD), the NLB disappears and the chromatin condenses to form an irregular network of individual bivalents (3). Despite the high similarity between nucleoli of GV oocytes and transcriptionally active embryonic cells, the condensed fibrillar spheres, i.e. NLBs in oocytes and NPBs in early embryos, exhibit slight differences. Tracing the material of oocyte NLBs material during the maturation and early post-fertilization stages shows that they are briefly existent in zygotes and early embryos, where the nucleolus is formed *de novo* in transcriptionally active embryonic cells (4). The key role of maternal nucleolus in embryonic development has been characterized in more detail, especially after the invention of a new method for microsurgical removal of NLBs (enucleolation). Its' reimplantation into oocytes and early embryos at different maturation or developmental stages has been reported with variable findings even between different animal species (5, 6, 7). These experiments unambiguously confirmed the importance of NLBs during normal development of mice and pig embryos, regardless of the method of their preparation (parthenogenetic activation, *in vitro* fertilization, or SCNT) (8, 9).

We hypothesized that NLBs and embryonic NPBs might support the embryonic development after parthenogenetic activation of oocytes previously subjected to interspecies nucleolar transfer. The current study aimed to elucidate the processes of rRNA during the development of interspecies (mouse/pig) nucleolus transferred embryos by reinjecting the NLBs from mouse or porcine GV stage oocytes into the previously enucleolated and matured porcine MII stage oocytes. Additionally, the study aimed to determine the embryonic development, timing, and intensity of *de novo* rRNA synthesis following the parthenogenetic activation of microsurgically manipulated versus non-manipulated porcine oocytes.

MATERIAL AND METHODS

Chemicals and media

Unless indicated otherwise, all chemicals were purchased from Sigma Chemical Co. (Prague, Czech Republic).

Ethical standards

All experiments in this research were conducted by the ethical standards of the relevant national regulations on the care and handling of farm and laboratory animals and the EU Directive 2010/63/EU on the protection of animals used for scientific purposes. The experimental protocol was approved by the Institutional Research Ethics Committee (permit no. 2547/19-312/4).

Collection of porcine oocytes

Porcine ovaries were collected at a local abattoir and transferred to the laboratory in saline buffer at 30-35 °C. After triple washing with PBS supplemented with 0.01% polyvinyl alcohol (PVA), the presumable GV-stage oocytes were aspirated from 3-6 mm follicles (10). The cumulus-oocyte complexes (COCs) were washed three times in M2 medium. COCs were *in vitro* matured in bicarbonate-buffered medium 199 supplemented with 4 mg/mL of bovine serum growth proteins, 0.5 µg/mL FSH, 0.5 µg/mL LH, 40 µg/mL sodium pyruvate, 70 µg/mL L-cysteine and 50 µg/mL gentamycin (Gibco Invitrogen, Prague, Czech Republic) (IVM) for 30 min at 38.5 °C and 5% CO₂ in humidified atmosphere. Subsequently, the cumulus cells were completely removed by gentle pipetting. Denuded oocytes were used for further enucleolation procedures.

Collection of mouse oocytes

Mouse ovaries were collected from 6–10-week-old ICR female mice (Anlab, Prague, Czech Republic), euthanized by a cervical dislocation. To obtain the GV stage oocytes, females were treated with 5 IU equine chorionic gonadotropin (eCG; Intervet International, Boxmeer, Holland) and after 44 hours, the GV oocytes were isolated from large antral follicles using a sharp needle. COCs were briefly pre-cultured in maturation medium (αMEM) supplemented with bovine serum albumin (BSA, 4 mg/mL), gentamicin (50 µg/mL), Na-pyruvate (20 mM) and dbcAMP (150 µg/mL) at 37 °C under an atmosphere of 5% CO₂. The oocytes were denuded from the surrounding follicular cells by vigorous pipetting.

Enucleolation of porcine and mouse GV oocytes

The porcine and mouse GV stage oocytes were enucleolated (ENL) according to a method described elsewhere (11). Briefly, GV stage oocytes were manipulated with a micromanipulator in M2 medium supplemented with 7.5 µg/mL cytochalasin B/D

and 0.1% PVA (Enu-M2). After brief centrifugation (5 min at 1,350 g), porcine oocytes were transferred into 10 μ L droplets of Enu-M2 covered with paraffin oil. For ENL, the enucleation pipette was inserted through the *zona pellucida* and slightly attached to the GV membrane surface. Subsequently, a gentle suction was applied on the pipette tip to translocate the nucleolus through the GV membrane. The enucleolated porcine oocytes were cultured until MII stage in IVM medium, as described above. The nucleoloplasts (nucleoli surrounded by plasma membrane) were temporarily kept in Enu-M2 medium before the re-injection into previously enucleolated oocytes.

Nucleolus re-injection and parthenogenetic activation

The enucleolated porcine MII oocytes were re-injected with recently isolated porcine (P+P) or mouse (P+M) nucleolus, as described previously (Fig. 1) (6). The nucleolus re-injected and intact (parthenogenetic control) porcine MII oocytes were cultured in porcine zygote medium-3 (PZM3) supplemented with 3 mg/mL BSA (PZM3-BSA) for 4 hours at 38.5 °C, 5% CO₂, and were then parthenogenetically activated. The oocytes were transferred into a 5.5% mannitol solution and activated by a single DC pulse of 1500 V/cm for

100 μ s (CF-150; BLS, Budapest, Hungary). The activated oocytes were washed at least three times in PZM3-BSA medium supplemented with 5 μ g/mL of cytochalasin B to prevent the exclusion of the second polar body. Cytochalasin B treated oocytes were washed in PZM3-BSA medium and cultured for 6 days. The embryos of experimental and control groups were collected at the zygote phase, 12 hours post activation (hpa), 2-cell (22 to 24 hpa), 4-cell (46 to 48 hpa), 8-cell (70 to 72 hpa), and blastocyst stage (140 to 144 hpa). They were processed for real-time qRT-PCR analysis.

Real-time qRT-PCR

The genes analyzed in the present study are crucial in the process of ribosome biogenesis organized in a nucleolus where the transcription, maturation, and assembly processes are performed.

In order to uncover the role of oocyte NLB on the initiation of rRNA transcription and processing, the levels of *de novo* synthesised 45S rRNA and processed 18S rRNA were observed during development of embryos acquired by parthenogenetic activation of intact (control) and nucleolus exchanged oocytes.

All the pools were done in triplicate and contained 3 embryos after parthenogenetic activation of nucleolus exchanged and intact oocytes, each at

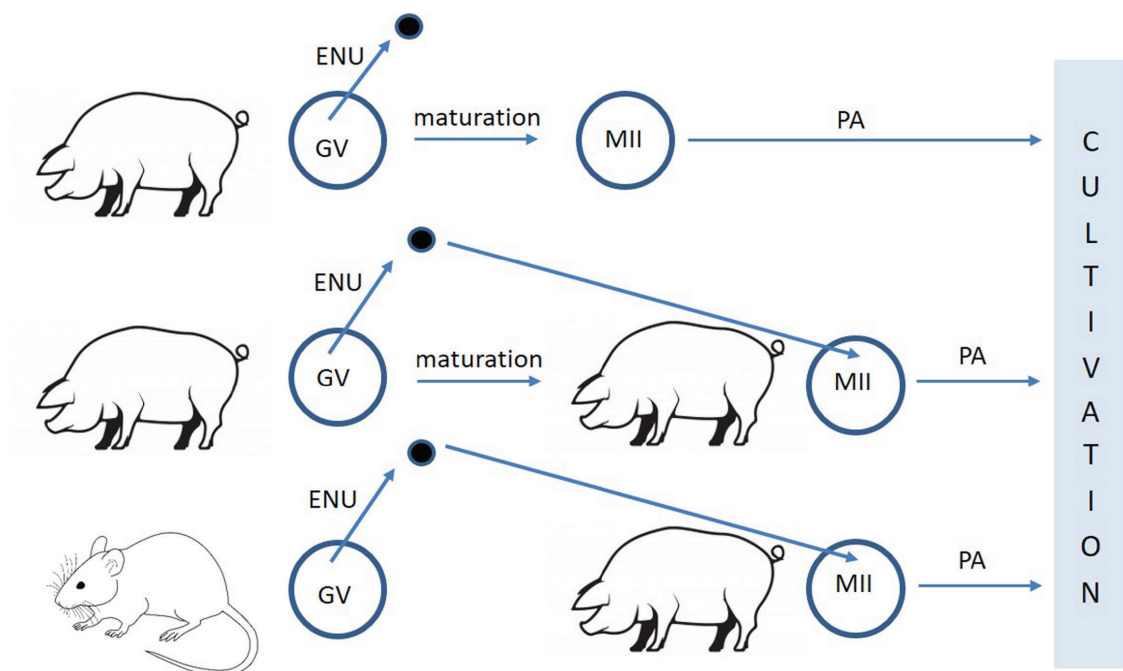


Figure 1. Experimental scheme of NLB micromanipulation procedure. Enu - enucleation; GV - germinal vesicle stage oocyte; MII - metaphase II stage oocyte; PA - parthenogenetic activation

Table 1. Porcine specific primer sequences designed for detection of *de novo* synthesised 45S rRNA and processed 18S rRNA

Primer	Sequence (5'-3')	Length
45S rRNA	F-AGACCTTGGGCCTCTGGAT, R-GCTGGATGCACAAGTGAGAG	134 bp
18S rRNA	F-TCGCGGAAGGATTAAAGTG, R-AAACGGCTACCACATCCAAG	141 bp

zygote, 2-cell, 4-cell, 8-cell, and blastocyst stages. Before RNA isolation, 2 pg of rabbit globin RNA (Gibco, Invitrogen, Carlsbad, CA) was added to the samples as an internal standard. The total RNA was extracted using the NucleoSpin RNA XS kit (Macherey-Nagel), and DNase I treatment was performed directly in the column as described by the manufacturer. Subsequently, reverse transcription was carried out using 2.5 µM random hexamers (Applied Biosystems, CA) and MuLV Reverse Transcriptase (Applied Biosystems, Foster City, CA) in total volume of 20 µL for 1 hour at 42 °C.

Real-Time PCR was performed on ABI 7500 Fast Real-Time System using Power SYBR® Green PCR Master Mix (Applied Biosystems, CA) containing MgCl₂, dNTP, and AmpliTaq Gold® DNA polymerase. Specific primer sequences for the detection of *de novo* synthesized 45S rRNA and processed 18S rRNA were designed as described in Table 1.

Statistics

All *in vitro* culture and qRT-PCR experiments were repeated more than three times. The maturation and developmental competence of oocytes and embryos in each group was analyzed by Fisher's exact test and the chi-square test. Mean values of qPCR data within each group (P+P, M+P, and parthenogenetic control) were analyzed by one-way analysis of variance method using SigmaStat 9.0 software (Systat Software Inc., San Jose, USA). Significant results from SigmaStat 9.0 were further analysed by Tukey test. Results from both experiments were considered significant at $p < 0.05$.

RESULTS

Developmental competence of nucleolus exchanged oocytes

ENL oocytes showed lower ($p < 0.05$, 50.6%, 37/73 oocytes) cleavage rate than the control (intact, 65.3%, 149/228 oocytes, Table 2). Furthermore, the blastocyst rate of the ENL rapidly decreased to 2.7% (2/73 oocytes) which was significantly lower ($p < 0.05$) than the control (30.7%, 70/228 oocytes) and experimental groups (11.1%, 17/153, and 13.8%, 23/166 oocytes, respectively) (Table 2).

After the re-injection of porcine NLBs into the porcine enucleolated MII stage oocytes, the reconstructed embryos partially regained the capability to reach the blastocyst stage (11.1%, 17/153 oocytes). The MII stage porcine oocytes re-injected with mouse NLB displayed very similar cleavage rate (56.6%, 94/166 oocytes) with the intraspecies group (58.8%, 90/153 oocytes). Moreover, 13.8% of these embryos reached to the blastocyst stage (Table 2).

Initiation of rRNA transcription and pre-rRNA processing

The synthesis of 45S rRNA and 18S rRNA was detected at the 8-cell stage in the control group (Fig. 2a, 2b). Further qPCR analyses of 45S rRNA synthesis rate showed that intra- and interspecies nucleolus re-injected embryos initiated lower *de novo* rRNA production at the 8-cell stage compared with the control (Fig. 2a). The processing of pre-rRNA in both experimental groups was initiated at blastocyst stage. Moreover, the porcine embryos

Table 2. The developmental potential of control, enucleolated and nucleolus re-injected porcine oocytes after parthenogenetic activation

Oocyte		No. of analysed oocytes	Developmental rates (%)	
Nucleoli	Cytoplasm		Cleaved (2 days)	Blastocysts (6 days)
Control (non-manipulated)		228	149 (65.3) ^a	70 (30.7) ^a
Enucleolated (ENL, non re-injection)		73	37 (50.6) ^b	2 (2.7) ^b
Pig	Pig	153	90 (58.8) ^{ab}	17 (11.1) ^c
Mouse	Pig	166	94 (56.6) ^b	23 (13.8) ^c

^{a,b,c}Values with different superscripts are significantly different ($p < 0.05$)

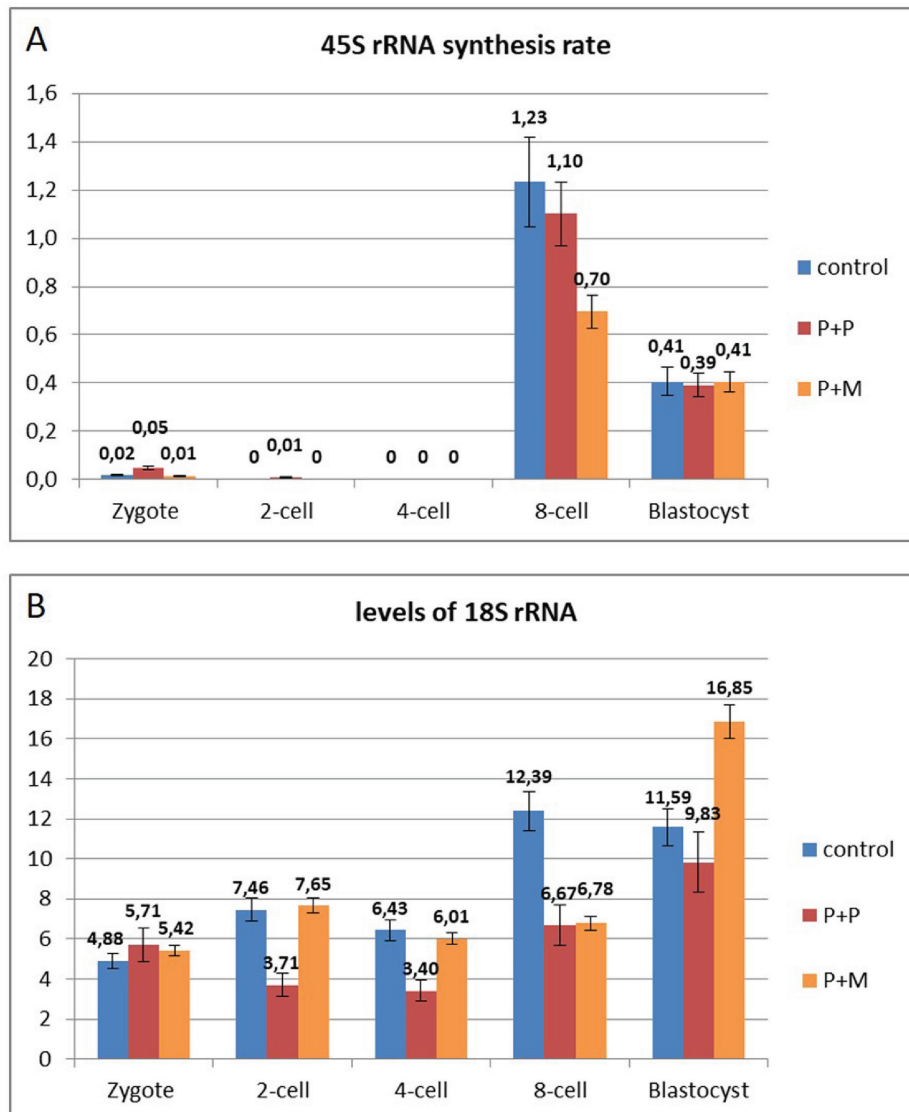


Figure 2. Relative expression of 45S rRNA (A) and levels of maternally inherited and *de novo* processed 18S rRNA (B) during development of porcine control (non-manipulated) and nucleolus re-injected intragenetic (P+P) and intergeneric (P+M) embryos estimated by RT-qPCR analysis. Mean values of ratios/levels are shown, with error bars representing the minimum and maximum measured values. Results of the rRNA expression and processing rate were statistically significant ($p < 0.05$, Tukey test)

with re-injected mouse NLB displayed significantly higher levels of 18S rRNA in the blastocyst stage compared with the control and experimental intraspecies groups (Fig. 2b).

DISCUSSION

The remodeling of fully functional nucleolus during major embryonic genome activation is closely related to the activation of rDNA transcription, pre-rRNA processing, and *de novo* synthesis

of ribosomes (2, 13). It is already known that NPBs are involved in the regulation of chromatin reorganization and formation of chromocenters with regulating potential for transcriptional silencing during early embryogenesis (9, 14). The importance of oocyte NLBs and embryonic NPBs in the above-mentioned processes was demonstrated in recent experiments where these structures were microsurgically removed from oocytes or zygotes and were subsequently re-injected at different time points of their maturation or development, using different animal species (mouse, pig) (15, 4).

In the present study, the enucleolation and nucleolus reinjection procedures were used to elucidate the importance of oocyte NLBs in development of porcine parthenogenetic embryos. Moreover, the study investigated the possible positive effects of interspecies nucleolar transfer on the developmental potential of formerly enucleolated porcine oocytes. The cleavage rate of parthenogenetically activated enucleolated porcine oocytes after the first two days was 50.6%. Only a fraction of the embryos reached the blastocyst stage (2.7%). Similar results were obtained in other studies where most of the fertilized or parthenogenetically activated mouse enucleolated oocytes, matured to MII phase, failed to cleave to the blastocyst stage. On the other hand, the developmental competence of mouse oocytes (MII) regained after the reinjection of NLBs from GV stage oocytes, with 50% reaching the blastocyst stage after fertilization (16). In contrast to studies on mouse oocytes, only 11.1% of the porcine oocytes re-injected with porcine NLB and 13.8% with mouse NLB, were able to reach to the blastocyst stage. This group of intraspecies NLB transferred embryos shows the adverse effect of the micromanipulation procedure accompanied with the potential loss of nucleolar material. To explain the low developmental capability of porcine nucleolus re-injected embryos found in this study, we decided to examine the exact timing of rDNA transcription and investigate the levels of unprocessed 45S rRNA and processed 18S rRNA during embryogenesis up to the blastocyst stage. qPCR analysis of 45S rRNA expression rate showed that the timing of genome activation in the embryos derived from porcine oocytes re-injected with porcine or mouse NLBs corresponds with the control group (8-cell stage), but the levels of transcripts are lower. These results also correspond with the timing of MGA in porcine parthenogenetic embryos detected in our previous study (12). These results are supported by experiments with mouse enucleolated oocytes re-injected with two porcine NLBs which were able to activate rRNA synthesis at the 4-cell stage, with slightly altered expression dynamics compared to the control (7). Interestingly, when the levels of 18S rRNA were measured, the experimental intra- and intergeneric groups of embryos showed delayed initiation of pre-rRNA processing at the blastocysts stage compared with the control. In alignment with the levels of 45S rRNA, these observations correspond with well-known theories about the exact balance between processed and unprocessed rRNA transcripts in mammalian cells (17). The

increased levels of 18S rRNA in blastocysts of intergeneric embryos compared with the control and intrageneric embryos, can be explained by higher protein amounts in mouse NLBs (1.55 ng) than those in pig (0.9 ng) (7). These mouse, protein-rich NLBs may contain higher levels of nucleolar proteins involved in the ribosome biogenesis (2) and thus provide more intensive support of pre-rRNA processing in porcine re-injected embryos.

CONCLUSION

The reinjection of NLBs from mouse GV oocytes into the enucleolated MII stage porcine oocytes can support embryonic development up to the blastocyst stage after its activation. Moreover, interspecies NLB transfer embryos initiated rDNA transcription at the same time as MGA in porcine parthenogenetic embryos. Despite the alterations in processing dynamics of these embryos, our results strongly point to the potential of applied NLB transfer technique in future studies focused on the improvement of mammalian oocyte quality and thus support the developmental potential of *in vitro* produced embryos.

CONFLICT OF INTEREST

The authors declare that they have no known conflict of interest in the conduction of the current study.

ACKNOWLEDGMENTS

This work was supported by the Slovak Research and Development Agency under the Contract no. DS-FR-22-0003 and by the projects “EXCELLENCE in molecular aspects of the early development of vertebrates”, CZ.02.1.01/0.0/0.0/15_003/0000460 from the Operational Programme Research, Development and Education and by the Danish Council for Independent Research/Natural Sciences (FNU) 8021-00048B. It was also co-funded by the projects VEGA 1/0167/20, 1/0270/24, KEGA 038UKF-4/2021, FVMS-IPR-02 (project no. 2020-2090/4).

AUTHORS' CONTRIBUTION

MM designed the study and concept of the analyses and wrote the manuscript. FS revised the manuscript. MB was involved in micromanipulation, data analysis and graphic design of experimental scheme. ARB contributed

in interpretation of qPCR data. SS involved in collection of mouse and porcine oocytes. FPP and MD revised the manuscript. JL gave a critical review and final approval of the version to be published. All authors have revised and approved the final version of the manuscript.

REFERENCES

1. Lavrentyeva, E., Shishova, K., Kagarlitsky, G., Zatsepina, O. (2015). Localisation of RNAs and proteins in nucleolar precursor bodies of early mouse embryos. *Reprod Fert Develop.* 29(3): 509-520. <https://doi.org/10.1071/RD15200>
2. Shishova, K.V., Lavrentyeva, E.A., Dobrucki, J.W., Zatsepina, O.V. (2015). Nucleolus-like bodies of fully-grown mouse oocytes contain key nucleolar proteins but are impoverished for rRNA. *Dev Biol.* 397(2): 267-281. <https://doi.org/10.1016/j.ydbio.2014.11.022>
3. Motlik, J., Fulka, J. (1986). Factors affecting meiotic competence in pig oocytes. *Theriogenology* 25(1): 87-96. [https://doi.org/10.1016/0093-691X\(86\)90185-8](https://doi.org/10.1016/0093-691X(86)90185-8)
4. Fulka, H., Aoki, F. (2016). Nucleolus precursor bodies and ribosome biogenesis in early mammalian embryos: old theories and new Discoveries. *Biol Reprod.* 94(6): 143. <https://doi.org/10.1095/biolreprod.115.136093>
5. Fulka, J.Jr, Moor, R.M., Loi, P., Fulka, J. (2003). Enucleation of porcine oocytes. *Theriogenology* 59(8): 1879-1885. [https://doi.org/10.1016/S0093-691X\(02\)01226-8](https://doi.org/10.1016/S0093-691X(02)01226-8)
6. Morovic, M., Strejcek, F., Nakagawa, S., Deshmukh, R.S., Murin, M., Benc, M., Fulka, H., et al. (2017). Mouse oocytes nucleoli rescue embryonic development of porcine enucleated oocytes. *Zygote* 25(6): 675-685. <https://doi.org/10.1017/S0967199417000491>
7. Benc, M., Martinkova, S., Rychtarova, J., Fulka, J.Jr., Bartkova, A., Fulka, H., Laurincik, J. (2020). Assessing the effect of interspecies oocyte nucleolar material dosage on embryonic development. *Theriogenology* 155, 17-24. <https://doi.org/10.1016/j.theriogenology.2020.06.001>
8. Ogushi, S., Palmieri, C., Fulka, H., Saitou, M., Miyano, T., Fulka, J.Jr. (2008). The maternal nucleolus is essential for early embryonic development in mammals. *Science* 319(5863): 613-616. <https://doi.org/10.1126/science.1151276>
9. Fulka, H., Langerova, A. (2014). The maternal nucleolus plays a key role in centromere satellite maintenance during the oocyte to embryo transition. *Development* 141(8): 1694-1704. <https://doi.org/10.1242/dev.105940>
10. Murin, M., Strejcek, F., Bartkova, A., Morovic, M., Benc, M., Prochazka, R., Lucas-Hahn, A., et al. (2019). Intranuclear characteristics of pig oocytes stained with brilliant cresyl blue and nucleologenesis of resulting embryos. *Zygote* 27(4): 232-240. <https://doi.org/10.1017/S0967199419000352>
11. Benc, M., Fulka, J.Jr., Strejcek, F., Morovic, M., Murin, M., Martinkova, S., Jettmarova, D., Laurincik, J. (2019). Enucleation and nucleolus transfer in mammalian oocytes and zygotes. *Int J Dev Biol.* 63(3-4-5): 253-258. <https://doi.org/10.1387/ijdb.190002mb>
12. Morovic, M., Østrup, O., Strejcek, F., Benc, M., Murin, M., Jedlickova, K., Bartkova, A., et al. (2018). Maternally inherited rRNA triggers de novo nucleolus formation in porcine embryos. *Zygote* 26(5): 395-402. <https://doi.org/10.1017/S0967199418000400>
13. Lafontaine, D.L.J., Riback, J.A., Bascetin, R., Brangwynne, C.P. (2021). The nucleolus as a multiphase liquid condensate. *Nat Rev Mol Cell Biol.* 22(3): 165-182. <https://doi.org/10.1038/s41580-020-0272-6>
14. Guthmann, M., Qian, C., Gialdini, I., Nakatani, T., Ettinger, A., Schauer, T., Torres-Padilla, M.E. (2023). A change in biophysical properties accompanies heterochromatin formation in mouse embryos. *Genes Dev.* 37(7-8): 336-350. <https://doi.org/10.1101/gad.350353.122>
15. Kyogoku, H., Fulka, J., Wakayama, T., Miyano, T. (2014). De novo formation of nucleoli in developing mouse embryos originating from enucleated zygotes. *Development* 141(11): 2255-2259. <https://doi.org/10.1242/dev.106948>
16. Ogushi, S., Saitou, M. (2010). The nucleolus in the mouse oocyte is required for the early step of both female and male pronucleus organization. *J Reprod Dev.* 56(5): 495-501. <https://doi.org/10.1262/jrd.09-184H>
17. Chakraborty, A., Uechi, T., Kenmochi, N. (2011). Guarding the „translation apparatus“: defective ribosome biogenesis and the p53 signaling pathway. *Wiley Interdiscip Rev RNA.* 2(4): 507-522. <https://doi.org/10.1002/wrna.73>

Please cite this article as: Morovic M., Strejcek F., Benc M., Rosenbaum Bartkova A., Stachovicova S., Popovska-Percinic F., Dovenska M., Laurincik J. Mouse nucleolus-like body supports transcription and enhances processing of rRNA in porcine enucleated oocytes. *Mac Vet Rev* 2025; 48 (1): 15-21. <https://doi.org/10.2478/macvetrev-2025-0011>