Macedonian Veterinary Review Mac Vet Rev 2026; 49 (2): i-xi

Available online at
www.macvetrev.mk

Original Scientific Article

INFLUENCE OF RELATIVE HUMIDITY ON INTERNAL QUALITY, BACTERIAL LOAD, AND MOLD CONTAMINATION OF EGGS DURING 28-DAY STORAGE

Ahmet Yavuz Pekel[1], Alp Emre Yıldız[2], Abdurrahman Kızıl[1], Ali Aydın[2]

[1]Department of Animal Nutrition and Nutritional Diseases, Faculty of Veterinary Medicine, Istanbul University-Cerrahpaşa, Istanbul 34320, Türkiye
[2]Department of Food Hygiene and Technology, Faculty of Veterinary Medicine, Istanbul University-Cerrahpaşa, Istanbul 34320, Türkiye

Received 12 March 2026; Received in revised form 4 June 2026; Accepted 11 July 2026

ABSTRACT

This study analyzed the effects of varying relative humidity (RH) on the internal and microbiological quality of eggs stored at ambient temperature (22±2°C) for 28 days. Medium-sized eggs, produced by 60-week-old Lohmann Brown hens (n=30), were selected randomly and allocated into sealed chambers set to 20%, 60%, or 100% RH. Egg weights were measured weekly, while internal quality attributes were evaluated at the conclusion of the storage duration. Eggs stored at 20% RH exhibited the highest weight loss (5.9%), whereas those at 100% RH showed minimal moisture loss (0.39%). Eggs stored at 20% RH also exhibited the highest yolk redness. Compared to 96.7% and 100% in the 60% and 20% RH groups, respectively, only 36.7% of eggs stored at 100% RH remained organoleptically acceptable for consumption after 28 days, based on predefined classification criteria. The 60% RH condition resulted in the poorest albumen structure, as reflected by the lowest Haugh unit scores. The length and width of the albumen decreased at 20% RH, while yolk pH remained unaffected by the humidity level. The yolk index declined as humidity increased. Eggs stored at 100% RH harbored the highest microbial loads, including mold, yeast, and aerobic bacteria, whereas the lowest shell contamination was observed at 20% RH. No significant differences in microbial counts were identified between the 20% and 60% RH groups. In conclusion, storage at low humidity (20%) was associated with better preservation of selected quality whereas saturated humidity (100%) was associated with significant quality deterioration and an increased risk of contamination.

Keywords: relative humidity, internal quality, eggshell microbial activity, egg color

INTRODUCTION

Eggs are highly regarded as a food product due to their rich nutrient profile and versatility. Nevertheless, their quality diminishes over time during storage, influenced by environmental factors such as temperature, relative humidity (RH), and duration of storage (1). Ensuring the maintenance of egg quality throughout storage is vital for safeguarding consumer acceptance and food safety (2). Egg quality encompasses both internal attributes (e.g., Haugh unit, yolk index, pH) and external characteristics (e.g., shell integrity, weight loss), all of which are substantially affected by storage conditions and duration (3). It is essential to determine suitable storage conditions to extend shelf life and maintain egg quality, benefiting both the food industry and consumers (4). A primary mechanism through which egg quality deteriorates during storage involves the diffusion of water and gases (such as CO_2) through the pores of the eggshell. As water evaporates and CO_2 escapes, the air cell enlarges, the albumen thins, and the pH elevates due to the loss of carbonic acid (5). These alterations can be quantified by parameters such as egg weight loss, Haugh unit (HU), and albumen height (6). The HU, first introduced by Haugh in 1937, remains one of the most prevalent

Corresponding author: Prof. Ahmet Yavuz Pekel, PhD
E-mail address: pekel@iuc.edu.tr
Present address: Department of Animal Nutrition and Nutritional Diseases, Faculty of Veterinary Medicine, Istanbul University-Cerrahpaşa, Istanbul 34320, Türkiye
Phone: +905322423347

Competing Interests: The authors have declared that no competing interests exist.
Available Online First: 27 July 2026
https://doi.org/10.2478/macvetrev-2026-0022

indicators of internal egg quality because it adjusts the height of the thick albumen relative to the egg's weight; higher HU values correspond to greater freshness and protein structural integrity (7). Prolonged storage, particularly under suboptimal environmental conditions, results in a reduction of HU, as demonstrated in studies involving refrigerated and room-temperature storage (8).

Relative humidity is a critical environmental variable with a dual function: it manages moisture exchange through the eggshell and influences the microbial environment present on the shell surface. These effects subsequently impact on both the physicochemical integrity and microbiological safety of eggs during storage (9). While low RH accelerates moisture loss from eggs - leading to weight reduction and decreased internal quality - excessively high RH may promote microbial proliferation on the shell surface, thereby risking the integrity of the shell and food safety (10). Kinetic modeling studies have illustrated that RH has a significant effect on both weight loss and HU, underscoring the importance of regulating RH during storage (11). Microbial contamination of the eggshell raises additional concerns, particularly when storage conditions favor mold and bacterial growth (12). The environments characterized by elevated humidity are linked to increased microbial loads on shell surfaces, which may penetrate the shell or contaminate eggs during handling (13). Despite the paramount importance of microbiological quality, numerous studies have predominantly concentrated on physicochemical and structural changes rather than on microbial dynamics across different RH levels (14). Extensive research has thoroughly documented how storage temperature and duration affect egg quality. However, few investigations have systematically evaluated how varying RH conditions - such as very low versus saturated environments - impact internal quality traits (including weight loss, albumen, yolk, and HU) and microbial contamination over extended storage periods. This significant knowledge gap is particularly relevant considering the considerable variability in storage environments (e.g., dry versus humid warehouses, regional disparities), where both quality and safety are imperative for consumer acceptance.

The study investigates the effects of storing eggs at three different RH levels (20%, 60%, and 100%) over 28 days, focusing on internal quality indicators such as egg weight loss, HU, yolk index, pH, and color, along with microbiological factors like aerobic bacteria, yeasts, and molds on the shell. The aim is to understand how RH influences the preservation of functional quality and the risk of microbial contamination, ultimately providing guidance for best practices in egg storage for both commercial and household settings.

MATERIAL AND METHODS

Fresh medium-sized eggs (n=30) were randomly collected from those laid by 60-week-old Lohmann Brown hens and stored for 28 days in an airtight vacuum oven (Nüve EV 018, Ankara, Türkiye) to maintain consistent RH. A total of 90 eggs were individually numbered and weighed, with 30 eggs assigned to each of the three treatment groups (20, 60, and 100% RH). No live animals were used in this study; therefore, ethical approval was not required. The study and data usage were performed with the informed consent and approval of the farm owner, who granted permission for the research and the reporting of the obtained data.

Relative humidity treatments

A 100% RH environment was established through the saturated water vapor methodology. In summary, a broad, shallow stainless-steel tray was filled with distilled water and positioned at the lower compartment of the vacuum oven. As air enclosed with liquid water at a constant temperature reaches saturation, this technique effectively creates an environment of 100% RH (15). The vacuum oven door was securely sealed, and the system was allowed to equilibrate for 2 h prior to placing the eggs. Relative humidity levels were continuously monitored with a digital hygrometer (CEM-DT-172, No. 11048007, Shenzhen, China) to ensure humidity remained within the range of 95% to 100% during the entire storage period.

The environment with 60% RH was established using a saturated potassium iodide (KI) solution, a standard method for generating intermediate humidity levels within sealed chambers. A saturated KI solution, containing excess solid crystals, was placed in an open glass tray at the bottom of the vacuum oven. According to the validated humidity reference data, saturated KI has a vapor pressure corresponding to approximately 69% RH at 25 °C, which generally stabilizes within the 60–65% RH range under standard incubator operating conditions (15). The chamber was allowed to equilibrate for 3 h before the introduction of eggs, with RH continuously monitored via a digital hygrometer.

A 20% RH environment was established utilizing activated silica gel as a desiccant. The silica gel underwent regeneration in an oven at 120–150 °C for 1–2 h prior to deployment, and was then placed in broad, shallow trays positioned around the egg racks on perforated supports to prevent direct contact with the eggs. A calibrated digital hygrometer was strategically positioned at the egg level to continuously monitor RH and temperature. The oven was allowed to reach equilibrium over 1-4 h, after which additional desiccant was added incrementally until the RH stabilized at 20%±5%. Desiccant trays were regenerated or replaced as necessary to sustain the target RH throughout the 28-day storage period. This methodology adhered to established desiccant-based humidity control standards (16, 17).

Egg quality measurements

Eggs were weighed using a precision scale (Sartorius TE3102S, Germany; with an accuracy of ±0.01 g). The percentage of weight loss was calculated by determining the difference between the initial and subsequent egg weights.

The diameter of the air cell within each egg was measured under candling light (Yensan Incubation Machines, Turkey) using a digital caliper (Mitutoyo, UK).

The pH of the yolk was determined utilizing a calibrated digital pH meter (HI83141, HANNA Instruments, USA). Each sample was measured at least twice, with a third measurement conducted in cases of inconsistency. The electrode underwent daily calibration using pH 7.0 and pH 9.0 buffer solutions, was rinsed with distilled water between measurements, and readings were documented upon stabilization.

After breaking the eggs onto a flat glass surface, measurements were performed on the raw albumen and yolk. The height of the albumen was measured at the midpoint between the yolk margin and the edge of the thick albumen employing the same tripod micrometer (18).

The height of the albumen was measured at the midpoint between the yolk margin and the thick albumen employing the same tripod micrometer (18). The length and width of the albumen were recorded on a flat glass surface using an electronic caliper.

The HU was computed utilizing albumen height (H, mm) and egg weight (W, g), following the formula established by Eisen et al. (19): HU=100×log (H−1.7W^{0.37}+7.6).

Color measurements

The evaluation of egg yolk coloration was performed utilizing a portable colorimeter (Model 3nh-NR100, Guangzhou, China). Before measurement, the device was calibrated using the standard white ceramic tile supplied by the manufacturer. Eggs were broken onto a glass surface, and the colorimeter aperture was precisely aligned with the surface of the yolk. Measurements were obtained at three separate locations on the yolk, with meticulous attention to avoid the yolk–albumen interface.

Color values were measured using the CIE Lab* system, where L* indicates lightness, a* represents the red–green axis, and b* corresponds to the yellow–blue axis. Measurements were performed under a D65 illuminant with a 10° standard observer. For each egg, the average of three readings was used for statistical analysis (20).

Microbiological analysis

The surface area of each egg shell (3×3 cm^2) was swabbed using a sterile cotton swab over a defined template, and the populations of total aerobic bacteria as well as yeast and mold were subsequently determined.

Microbiological analyses were conducted at the beginning of the storage period and after 17 days of storage at ambient temperature (22±2 °C). On day 0, no statistically significant differences in eggshell microbiological counts were observed among the different treatment groups ($p>0.05$).

Owing to concurrent textural analyses, the reference method ISO 4833-1:2013 for total mesophilic aerobic bacteria was modified (21). Swab samples collected from a 3×3 cm^2 eggshell area were inoculated onto Plate Count Agar (PCA, Merck) and incubated at 37 °C for 24-48 h prior to enumeration.

Similarly, the reference method ISO 21527-1:2008 for yeast and mold enumeration was adapted (22). Swab samples were inoculated onto Yeast Glucose Chloramphenicol Agar (YGC Agar, Merck) and incubated at 25 °C for 5 days. Final results were expressed as colony-forming units per egg (CFU/egg).

All microbiological analyses were conducted in triplicate, and the results were transformed into logarithmic values (log CFU/egg) prior to statistical analysis.

Statistical analysis

All statistical analyses were conducted utilizing IBM SPSS Statistics (Version 26.0; IBM Corp., Armonk, NY, USA).

Each egg was designated as the experimental unit (n=30 per treatment). The normality of the data was evaluated via the Shapiro–Wilk test and Q–Q plots, whereas the homogeneity of variances was assessed through Levene's test.

Repeated-measures ANOVA was employed to analyze variables measured across multiple time points, with treatment serving as the between-subject factor and storage duration as the within-subject factor. Mauchly's test of sphericity was conducted, and Greenhouse–Geisser corrections were applied as appropriate. Post hoc comparisons were executed using Bonferroni adjustments.

Single-time-point variables were analyzed using one-way ANOVA with Tukey's post hoc test or Welch's ANOVA with Games–Howell correction when variance homogeneity was violated. Nonparametric data were analyzed using the Kruskal–Wallis test followed by Dunn's post hoc test with Bonferroni correction.

Categorical outcomes were assessed utilizing the Chi-square test or Fisher's exact test, as appropriate. Pearson or Spearman correlation analyses were conducted contingent upon the data distribution. All statistical tests were two-sided, with a significance threshold established at $p<0.05$. The results are displayed as mean ± standard error of the mean (SEM).

A multiple linear regression analysis was conducted to identify the predictors of the HU. Upon identifying multicollinearity within the comprehensive model, a stepwise elimination method was applied, yielding an optimized model that included albumen height, albumen width, yolk height, yolk index, and initial egg weight. The validity of this model was confirmed by a high coefficient of determination ($R^2=0.966$).

RESULTS

Egg weight

The impact of RH on egg weight over a 28-day storage period is presented in Table 1. Initial egg weights showed no significant differences among the treatment groups ($p>0.05$). Likewise, no statistically significant differences were observed at day 7 ($p>0.05$); however, from day 14 onwards, RH significantly influenced egg weight ($p<0.001$). Eggs stored at 100% RH retained substantially higher weights at days 14, 21, and 28 compared to those stored at 60% or 20% RH. By day 28, eggs stored at 100% RH had an average weight of 59.0 grams, whereas those stored at 60% and 20% RH decreased to 56.7 g and 56.3 g, respectively.

Table 1. The effect of different relative humidity levels on egg weight during 28-day storage

	Storage time (day)				
Relative humidity (%)	**0**	**7**	**14**	**21**	**28**
100	59.30±1.71	59.20±1.74	59.20^{a}±1.73	59.10^{a}±1.72	59.00^{a}±1.72
60	59.70±1.55	58.80±1.54	57.90^{b}±1.56	57.20^{b}±1.58	56.70^{b}±1.61
20	59.80±1.75	58.50±1.74	57.50^{b}±1.75	56.90^{b}±1.75	56.30^{b}±1.76
SEM	0.176	0.177	0.191	0.204	0.218
p-value	NS	NS	<0.001	<0.001	<0.001

N=30 per treatment
*NS: not significant

Table 2. The effect of different relative humidity levels on percentage egg weight loss during 28-day storage

	Storage time (day)			
Relative humidity (%)	**7**	**14**	**21**	**28**
100	0.05^{c}±0.27	0.10^{c}±0.23	0.20^{c}±0.22	0.39^{c}±0.39
60	1.60^{b}±0.35	3.10^{b}±0.70	4.10^{b}±0.93	5.00^{b}±1.14
20	2.20^{a}±0.38	3.80^{a}±0.66	4.90^{a}±0.84	5.90^{a}±1.00
SEM	0.101	0.180	0.231	0.274
p-value	<0.001	<0.001	<0.001	<0.001

N=30 per treatment

Percentage egg weight loss

As demonstrated in Table 2, RH markedly influenced egg weight loss throughout the storage duration ($p<0.001$ at all-time points). Eggs stored at 100% RH displayed the minimal weight loss during storage, with a cumulative loss of merely 0.39% by day 28. Conversely, eggs stored at 20% RH experienced the greatest weight loss at each assessment point, reaching 5.9% by day 28. Eggs stored at 60% RH exhibited intermediate weight loss values between these two extremes.

Egg condition after 28 days of storage

Egg condition classifications after 28 days of storage are summarized in Table 3. Relative humidity significantly influenced the distribution of normal, decomposed, and spoiled eggs ($p<0.001$). The highest proportion of normal eggs was observed at 20% RH (100%), followed by 60% RH (96.7%). Eggs stored at 100% RH showed the lowest proportion of normal eggs (36.7%) and the highest proportions of decomposed (40%) and spoiled eggs (23.3%). No spoiled eggs were recorded in the 60% or 20% RH treatment groups.

Air cell diameter and albumen quality

The effects of RH on air cell diameter and albumen characteristics after 28 days of storage are presented in Table 4. Air cell diameter differed significantly among treatments ($p<0.001$). Eggs stored at 100% RH exhibited the smallest air cells (18.8 mm), whereas eggs stored at 60% and 20% RH showed significantly larger air cell diameters (25.3 mm and 25.9 mm, respectively).

Albumen length, width, and height were also significantly affected by humidity ($p<0.001$). Eggs stored at 100% RH showed the highest albumen height (5.83 mm) as well as the greatest albumen length and width. In contrast, eggs stored at 20% RH exhibited marked reductions in albumen dimensions. Haugh unit values differed significantly among treatments ($p<0.001$), with the lowest value observed at 60% RH (56.76). Eggs stored at 100% and 20% RH maintained significantly higher HU values (71.35 and 72.29, respectively), indicating better albumen quality under both very high and very low humidity conditions.

Table 3. The effect of different relative humidity levels on egg condition after 28 days of storage

	Egg classification (%)		
Relative humidity (%)	**Normal**	**Decomposed**	**Spoiled**
100	$36.70^{b}\pm30.55$	$40.00^{a}\pm26.46$	$23.30^{a}\pm15.28$
60	$96.70^{a}\pm5.77$	$3.30^{b}\pm5.77$	$0.00^{b}\pm0.00$
20	$100.00^{a}\pm0.00$	$0.00^{b}\pm0.00$	$0.00^{b}\pm0.00$
SEM	0.04	0.04	0.03
p-value	<0.001	<0.001	<0.001

N=30 per treatment

Table 4. The effect of different relative humidity levels on air cell diameter, albumen properties, and Haugh unit after 28 days of storage

Relative humidity (%)	Air cell diameter	Albumen width	Albumen length	Albumen height	Haugh unit
100	$18.80^{b}\pm2.72$	$141.40^{a}\pm51.39$	$199.12^{a}\pm49.35$	$5.83^{a}\pm2.34$	$71.35^{a}\pm22.21$
60	$25.30^{a}\pm2.65$	$138.90^{a}\pm25.04$	$171.35^{b}\pm37.27$	$3.80^{b}\pm1.14$	$56.76^{b}\pm14.33$
20	$25.90^{a}\pm2.50$	$94.70^{b}\pm10.49$	$115.71^{c}\pm12.27$	$5.27^{a}\pm0.65$	$72.29^{a}\pm5.69$
SEM	0.440	4.155	5.435	0.178	1.815
p-value	<0.001	<0.001	<0.001	<0.001	<0.001

N=30 per treatment

Table 5. The effect of different relative humidity levels on egg yolk quality traits after 28 days of storage

Relative humidity (%)	Diameter	Height	Yolk index	L*	a*	b*	pH
100	42.52±1.25	13.10±4.80	26.80^{c}±3.68	18.30±1.41	8.33^{ab}±0.38	26.20±1.61	6.40±0.31
60	41.81±2.42	13.20±0.96	32.28^{a}±2.41	17.20±2.35	7.99^{b}±0.74	25.20±2.13	6.50±0.74
20	42.54±1.41	13.90±0.82	30.11^{b}±2.35	17.90±1.24	8.63^{a}±0.96	26.50±2.33	6.70±0.44
SEM	0.228	0.254	0.386	0.214	0.097	0.255	0.070
p-value	NS	NS	<0.001	NS	<0.05	NS	NS

N=30 per treatment; *NS: not significant

Table 6. Effect of different relative humidity levels on total aerobic bacteria and mold-yeast counts on eggshells after 17 days of storage (log_{10}cfu/cm^2)

Relative humidity %	Total aerobic bacteria	Mold-Yeast
100	3.31^{a}±0.07	3.19^{a}±0.09
60	1.42^{b}±0.53	1.76^{b}±0.52
20	2.02^{b}±0.67	1.40^{b}±0.40
SEM	0.171	0.102
p-value	<0.05	<0.05

N=30 per treatment

Yolk quality traits

The quality characteristics of the yolk evaluated after a 28-day storage period are detailed in Table 5. Humidity levels did not exert a statistically significant influence on yolk diameter, yolk height, or yolk pH ($p>0.05$). Nevertheless, the yolk index displayed significant variation among treatment groups ($p<0.001$), with the lowest observed at 100% RH and the highest at 60% RH. The yolk color parameter a* (redness) was also significantly affected by humidity ($p<0.05$), with eggs stored at 20% RH exhibiting the highest redness values. Other color parameters, namely L* and b*, were not significantly influenced by humidity levels ($p>0.05$).

Microbiological counts on eggshells

The effects of RH on total aerobic bacteria and mold–yeast counts on eggshells after 17 days of storage are presented in Table 6. Relative humidity markedly influenced both microbial parameters ($p<0.05$). Eggs stored at 100% RH demonstrated the highest total aerobic bacterial load (3.31 log_{10} CFU/cm^2) and mold–yeast counts (3.19 log_{10} CFU/cm^2). Conversely, eggs stored at 60% and 20% RH exhibited significantly reduced microbial loads, with no statistically significant difference observed between these two conditions.

Correlation analysis

The correlation analysis of egg quality parameters measured after 28 days of storage is depicted in Fig. 1. Robust positive correlations were observed among percentage weight-loss values across various storage intervals. The correlation between percentage weight loss at 0-14 days and 0-21 days was nearly perfect (r=0.9982). Likewise, percentage weight loss from 0-21 days showed a strong correlation with that from 0-28 days (r=0.9982). A significant correlation was also observed between percentage weight loss at 0-14 days and 0-28 days (r=0.9947). Furthermore, egg weights recorded at days 21 and 28 demonstrated a high degree of correlation (r=0.9939).

Relative humidity exhibited significant negative correlations with the percentage of weight loss across all storage durations, including 0-7 days (r=-0.9002), 0-14 days (r=-0.8916), 0-21 days (r=-0.8774), and 0-28 days (r=-0.8715). Furthermore, RH was significantly negatively correlated with air cell diameter (r=-0.7058), indicating reduced moisture loss and air cell expansion under elevated humidity.

Multiple regression analysis

A simplified multiple linear regression model was developed to identify the most significant predictors of the HU (refer to Table 7). The reduced

model demonstrated a high level of statistical significance (R^2=0.966; p<0.001). Albumen height was identified as the most influential predictor of the HU (p<0.001), followed by albumen width (p=0.002), initial egg weight (p=0.014), yolk height (p=0.024), and yolk index (p=0.046). All regression coefficients were positive, indicating that increases in these parameters are associated with higher HU values. Although multicollinearity affected the overall regression model, the simplified model effectively highlighted the variables most relevant for predicting albumen quality post-storage.

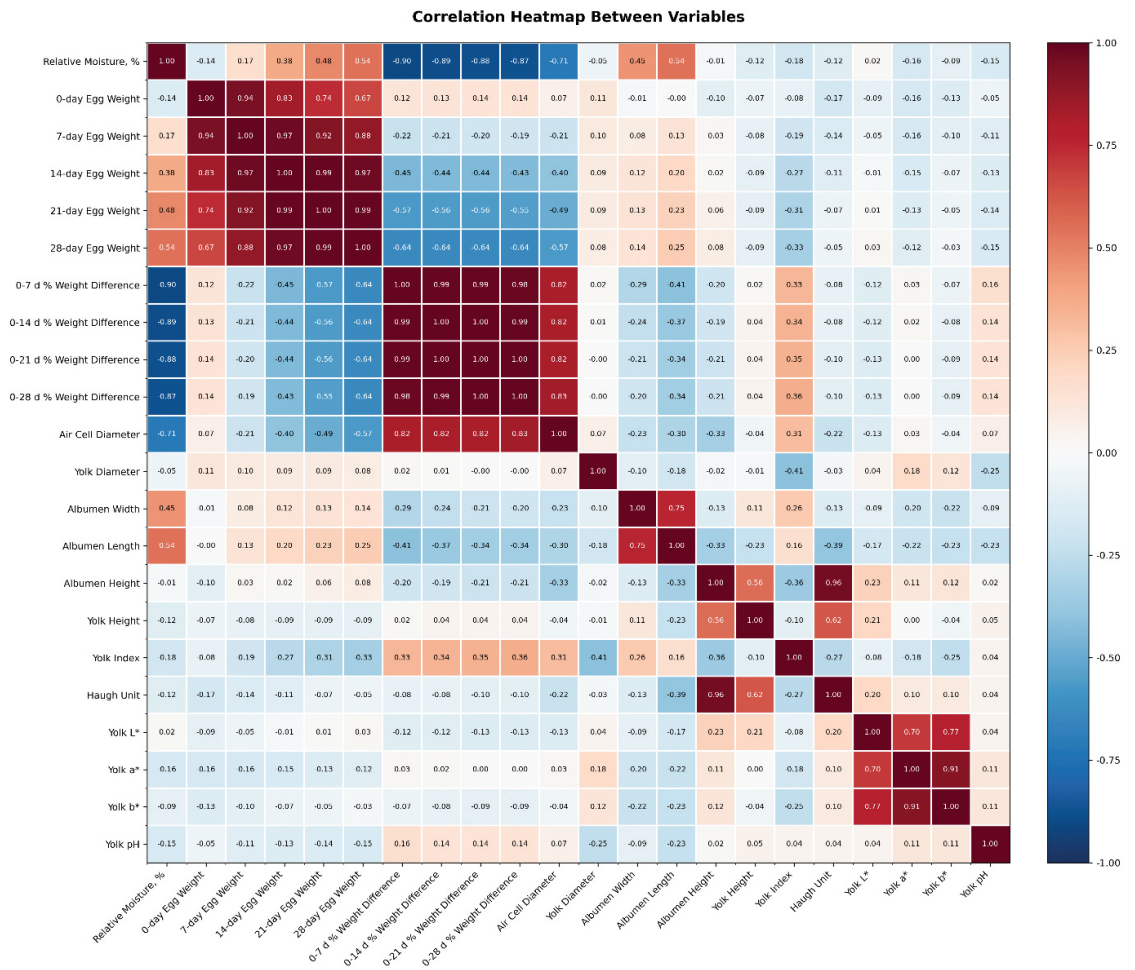

Figure 1. Correlation heatmap of interior egg quality parameters after 28 days of storage under different relative humidity levels

Table 7. Simplified multiple linear regression model for predicting Haugh unit in stored eggs

Predictor variable	Coefficient (β)	p-value	Significance
Intercept	10.1580	—	—
Albumen height (mm)	9.4826	0.000	***
Albumen width (mm)	0.0698	0.002	**
0-day egg weight (g)	0.2617	0.014	*
Yolk height (mm)	0.5808	0.024	*
Yolk index	0.3460	0.046	*

Model statistics: R^2=0.966, Overall model p<0.001
Severe multicollinearity detected in full model; simplified model retains only significant predictors.
Significance codes: ***p<0.001; **p<0.01; *p<0.05
Haugh unit=10.1580+0.2616869×(0 day egg weight)+0.0698×(Albumen width)+9.4826×(Albumen height)+0.5808×(Yolk height) +0.3460×(Yolk index)

DISCUSSION

This study systematically examined the effects of three different RH levels (20, 60, and 100%) on the internal quality and microbiological safety of eggs stored at room temperature for 28 days. The results show that RH has a complex dual role in egg preservation, affecting both physicochemical degradation and the likelihood of microbial contamination.

Impact of relative humidity on egg weight loss and internal quality

As anticipated, the greatest egg weight loss (5.9%) was recorded at the lowest RH level (20%), whereas the lowest loss (0.39%) occurred at 100% RH. This observation is consistent with the well-established principle that water vapor loss through the eggshell is driven by the vapor pressure gradient between the egg interior and the surrounding environment (5). Under low RH conditions, this gradient becomes steeper, accelerating moisture loss and leading to air cell enlargement. Conversely, high RH reduces the vapor pressure deficit, thereby limiting water loss and maintaining egg weight.

Nonetheless, the relationship between weight reduction and internal quality parameters was not strictly linear. The most significant decline in albumen quality was observed at 60% RH, where both albumen height and HU reached their minimum values. Generally, excessive moisture and carbon dioxide (CO_2) loss during storage are acknowledged to promote albumen thinning and a reduction in HU (10, 23). Interestingly, eggs stored at 20% RH retained comparatively high HU values despite experiencing considerable weight loss. This indicates that, under extremely dry conditions, water loss may occur more rapidly than CO_2 diffusion, which is regarded as a primary factor in albumen alkalization and thinning (3, 10). Alternatively, very low RH levels may stabilize albumen protein interactions, particularly between ovomucin and lysozyme, thereby retarding structural deterioration (24).

In contrast, the intermediate RH condition (60%) was the least effective in maintaining albumen quality, suggesting that moderate humidity levels may promote CO_2 diffusion relative to water loss, thereby resulting in a deterioration of the albumen structure. This observation underscores the intricate interaction between moisture loss, gas exchange mechanisms, and protein stability throughout egg storage (25, 26). The lower HU observed at 60% RH compared to 20% RH may be explained by the specific interaction between moisture loss and gas exchange mechanisms. At the intermediate level of 60% RH, the moderate vapor pressure deficit likely promotes a more efficient diffusion of CO_2 relative to water loss, which accelerates albumen alkalization and the subsequent thinning of the thick albumen (10, 23). Conversely, the rapid dehydration occurring at 20% RH may induce a "concentration effect" within the albumen matrix, physically stabilizing the interaction between ovomucin and lysozyme and potentially retarding the rate of CO_2 diffusion through the shell membranes (3, 24). This suggests that while 60% RH facilitates the biochemical processes leading to albumen deterioration, the extreme dryness at 20% RH creates a physical environment that paradoxically helps maintain the structural integrity of the albumen despite significant weight loss (25, 26). The relatively poor albumen quality observed at 60% RH indicates that intermediate humidity levels may not provide optimal conditions for preserving internal egg quality. Further studies are warranted to clarify the mechanisms governing CO_2 diffusion, moisture dynamics, and protein stability under moderate humidity conditions.

Morphological albumen measurements further supported these observations. Eggs stored at 20% RH exhibited significantly reduced albumen length and width, indicating a more compact and less hydrated albumen matrix. This structural compactness may contribute to maintaining albumen height and HU despite high overall moisture loss. Yolk traits were generally less sensitive to RH than albumen-related characteristics. However, yolk redness (a*) was significantly higher in eggs stored at 20% RH, likely due to pigment concentration effects of carotenoids associated with dehydration. Meanwhile, the yolk index was lowest at 100% RH, suggesting a weakening of the vitelline membrane under highly humid conditions, likely due to increased water migration from the albumen to the yolk driven by osmotic pressure (27, 28, 29). Notably, yolk pH remained unaffected by RH, indicating that the dynamics of CO_2 exchange may vary between egg compartments or that the impact of RH on yolk pH is comparatively limited in relation to its influence on albumen quality. Comparable findings have been documented in previous research examining gas exchange and internal egg chemistry during storage (30).

Microbiological safety and edibility

From a food safety standpoint, the most noteworthy discovery was the considerable decrease in the quantity of eggs suitable for consumption at 100% RH. Only 37% of eggs stored under saturated humidity conditions remained fit for consumption

after 28 days, in contrast to 97–100% in the groups maintained at 60% and 20% RH. This finding supports the hypothesis that elevated humidity levels promote conditions favorable to microbial proliferation on the eggshell surface (12).

Microbiological analyses confirmed that eggs stored at 100% RH demonstrated significantly higher levels of aerobic bacteria and molds/yeasts. In alignment with previous research, increased humidity promotes fungal growth and, in some cases, penetration into the egg's interior, thereby leading to spoilage or mycotoxin formation (12, 31). Moist shell surfaces enhance microbial adhesion, germination, and proliferation, and may also facilitate microbial penetration through shell pores or microcracks (32).

Conversely, eggs stored at 20% RH showed minimal microbial growth, supporting the hypothesis that a dry shell environment inhibits microbial proliferation. The absence of significant differences in microbial counts between the 20% and 60% RH groups indicates that moderate humidity levels do not markedly increase contamination risk within the examined storage duration and conditions. In contrast, saturated humidity distinctly enhances food safety risks. These findings indicate that minimizing weight loss alone should not be considered an adequate indicator of successful egg storage, as preservation of microbiological safety is equally essential.

Although maintaining a RH of 100% effectively prevents moisture loss, it leads to unacceptable microbiological deterioration. Consequently, the regulation of RH during egg storage must strike a balance between conserving physicochemical properties and controlling microbial proliferation.

Interrelationships of quality parameters

Correlation analysis revealed significant inverse relationships between RH and percentage weight loss across all storage intervals, thereby confirming that higher humidity reliably reduces moisture loss. Additionally, a notable negative correlation was observed between RH and air cell diameter, further corroborating the impact of environmental moisture on evaporation.

Strong positive correlations were observed between albumen height and HU, reaffirming the well-established significance of albumen structure as the primary determinant of internal egg quality (33). These relationships were further corroborated through the simplified multiple linear regression analysis, where albumen height was identified as the most influential predictor of HU, followed by albumen width, yolk height, yolk index, and initial egg weight. Similar findings have been documented in both chicken and quail eggs, emphasizing the robustness of albumen height as an indicator of freshness across various species and storage conditions (34, 35).

The inclusion of initial egg weight as a predictor suggests that the original egg size may influence the rate of quality deterioration during storage, potentially owing to differences in albumen-to-yolk ratios or shell characteristics (19). The final regression model was biologically plausible and demonstrated significant explanatory power, offering a practical approach for predicting internal egg quality based on a limited set of measurable parameters. Future research should investigate the combined effects of RH, storage temperature, packaging systems, and shell surface treatments to further optimize egg quality preservation and microbiological safety during prolonged storage.

CONCLUSION

In conclusion, RH significantly influenced both the internal quality and microbiological safety of eggs during 28 days of storage at room temperature. Storage at 20% RH provided the best preservation of egg quality, maintaining higher HU values and lower microbial contamination despite greater weight loss. In contrast, storage at 100% RH minimized weight loss but resulted in increased bacterial and mold contamination and a higher incidence of spoiled eggs. These findings indicate that preserving egg quality requires consideration of both internal quality and microbiological safety, and that low-humidity storage (approximately 20% RH) provides the most favorable conditions for prolonged egg storage.

CONFLICT OF INTEREST

The authors declare that they have no financial or non-financial conflict of interest regarding authorship and publication of this article.

ACKNOWLEDGMENTS

The authors gratefully acknowledge the staff of the Poultry Unit, Faculty of Veterinary Medicine, Istanbul University-Cerrahpaşa, Türkiye, for their valuable assistance with egg collection and handling throughout the study. This research received no external funding.

AUTHORS' CONTRIBUTION

AYP and AEY conceptualized the study, carried out and supervised the sample analysis, prepared graphs and tables, and wrote the manuscript. AK and AA were included in the hypothesis formulation. AA participated in editing the manuscript and contributed to the practical performance of this research. All authors have revised and approved the final version of the manuscript.

REFERENCES

1. Jones, D.R., Musgrove, M.T. (2005). Effects of extended storage on egg quality factors. Poult Sci. 84(11): 1774-1777. https://doi.org/10.1093/ps/84.11.1774 PMid:16463976
2. Kulshreshtha, G., Ward, C., Calvert, N.D., Benavides-Reyes, C., Rodriguez-Navarro, A.B., Diep, T., Hincke, M.T. (2024). Effect of egg washing and hen age on cuticle quality and bacterial adherence in table eggs. Microorganisms. 12(10): 2027. https://doi.org/10.3390/microorganisms12102027 PMid:39458336 PMCid:PMC11509795
3. Samli, H.E., Agma, A., Senkoylu, N. (2005). Effects of storage time and temperature on egg quality in old laying hens. J Appl Poult Res. 14(3): 548-553. https://doi.org/10.1093/japr/14.3.548
4. Cortés-Ramírez, G.S., Velasco, J.I., Plascencia, M.Á., Absalón, Á.E., Cortés-Espinosa, D.V. (2024). Development of edible coatings based on different biopolymers to enhance the internal shelf-life quality of table eggs. Coatings. 14(12): 1525. https://doi.org/10.3390/coatings14121525
5. Cozzolino, D., Sanal, P., Soumeh, E.A., Dekkers, M.H., Anderson, M., Boisen, S., Hoffman, L.C. (2024). Implications of the storage conditions and production system on the physical properties of eggs: a multivariate data analysis study. Int J Food Sci Technol. 59(5): 3235-3244. https://doi.org/10.1111/ijfs.17069
6. Quan, C., Xi, Q., Shi, X., Han, R., Du, Q., Forghani, F., Wang, J. (2021). Development of predictive models for egg freshness and shelf-life under different storage temperatures. Food Qual Saf. 5, fyab021. https://doi.org/10.1093/fqsafe/fyab021
7. Jones, D. (2012). Haugh unit: gold standard of egg quality. Natl Egg Qual Sch Proc. 7, 47-51.
8. Ikusika, O.O., Dwakasa, H., Luphuzi, S., Akinmoladun, O.F., Mpendulo, C.T. (2025). Assessment of egg quality across seasons, storage durations, and temperatures in commercial laying hens. Appl Sci. 15(19): 10344. https://doi.org/10.3390/app151910344
9. Wengerska, K., Batkowska, J., Drabik, K. (2023). The eggshell defect as a factor affecting the egg quality after storage. Poult Sci. 102(7): 102749. https://doi.org/10.1016/j.psj.2023.102749 PMid:37220686 PMCid:PMC10213368
10. Scott, T.A., Silversides, F.G. (2000). The effect of storage and strain of hen on egg quality. Poult Sci. 79(12): 1725-1729. https://doi.org/10.1093/ps/79.12.1725 PMid:11194033 PMCid:PMC8258603
11. Yimenu, S.M., Koo, J., Kim, J.Y., Kim, J.H., Kim, B.S. (2018). Kinetic modeling impacts of relative humidity, storage temperature, and air flow velocity on various indices of hen egg freshness. Poult Sci. 97(12): 4384-4391. https://doi.org/10.3382/ps/pey334 PMid:30085286 PMCid:PMC6305835
12. Tomczyk, Ł., Szablewski, T., Stuper-Szablewska, K., Nowaczewski,S., Cegielska-Radziejewska, R. (2019). The influence of the conditions of acquisition and storage of table eggs on changes in their quality and the presence of mycobiota and Fusarium mycotoxins. Poult Sci. 98(7): 2964-2971. https://doi.org/10.3382/ps/pez156 PMid:30951601
13. De Reu, K., Grijspeerdt, K., Heyndrickx, M., Messens, W., Uyttendaele, M., Debevere, J., Herman, L. (2006). Influence of eggshell condensation on the bacterial eggshell penetration and the whole egg contamination with Salmonella enterica serovar Enteritidis. J Food Prot. 69(7): 1539-1545. https://doi.org/10.4315/0362-028X-69.7.1539 PMid:16865883
14. Sokołowicz, Z., Kačániová, M., Dykiel, M., Augustyńska-Prejsnar, A., Topczewska, J. (2023). Influence of storage packaging type on the microbiological and sensory quality of free-range table eggs. Animals (Basel). 13(12): 1899. https://doi.org/10.3390/ani13121899 PMid:37370410 PMCid:PMC10295111
15. Greenspan, L. (1977). Humidity fixed points of binary saturated aqueous solutions. J Res Natl Bur Stand - A Phys Chem. 81A(1): 89-96. https://doi.org/10.6028/jres.081A.011 PMCid:PMC5295834
16. ASTM (1985). Standard practice for maintaining constant relative humidity by means of aqueous solutions. ASTM E104-85 (pp. 790-795). Philadelphia, PA: ASTM International
17. Young, J.F. (1967). Humidity control in the laboratory using salt solutions-a review. J Appl Chem. 17(9): 241-245. https://doi.org/10.1002/jctb.5010170901

18. Roberts, J.R. (2005). Egg quality guidelines for the Australian egg industry. Australian Egg Corporation Limited, North Sydney, Australia

19. Eisen, E.J., Bohren, B.B., McKean, H.E. (1962). The Haugh unit as a measure of egg albumen quality. Poult Sci. 41(5): 1461-1468. https://doi.org/10.3382/ps.0411461

20. Ortiz, D., Lawson, T., Jarrett, R., Ring, A., Scoles, K.L., Hoverman, L., Rocheford, T. (2021). Biofortified orange corn increases xanthophyll density and yolk pigmentation in egg yolks from laying hens. Poult. Sci. 100(7): 101117. https://doi.org/10.1016/j.psj.2021.101117 PMid:34102484 PMCid:PMC8187250

21. ISO. (2013). Microbiology of the food chain - Horizontal method for the enumeration of microorganisms - Part 1: Colony count at 30 °C by the pour plate technique. ISO 4833-1:2013. International Organization for Standardization, Geneva, Switzerland

22. ISO. (2008). Microbiology of food and animal feeding stuffs - Horizontal method for the enumeration of yeasts and moulds - Part 1: Colony count technique in products with water activity greater than 0.95. ISO 21527-1:2008. International Organization for Standardization, Geneva, Switzerland

23. Sheng, L., Huang, M., Wang, J., Xu, Q., Hammad, H.H.M., Ma, M. (2018). A study of storage impact on ovalbumin structure of chicken egg. J Food Eng. 219, 1-7. https://doi.org/10.1016/j.jfoodeng.2017.08.028

24. Hassan, A., Aylin, A.O. (2009). Effect of storage time and temperature on egg quality. J Anim Vet Adv. 8, 133-137.

25. Vlčková, J., Tůmová, E., Míková, K., Englmaierová, M., Okrouhlá, M., Chodová, D. (2019). Changes in the quality of eggs during storage depending on the housing system and the age of hens. Poult Sci. 98(11): 6187-6193. https://doi.org/10.3382/ps/pez401 PMid:31287891

26. Hidalgo, A., Lucisano, M., Comelli, E.M., Pompei, C. (1996). Evolution of chemical and physical yolk characteristics during the storage of shell eggs. J Agric Food Chem. 44(6): 1447-1452. https://doi.org/10.1021/jf950591q

27. Al-Hajo, N.N., Zangana, B.S., Al-Janabi, L.A., Al-Khalani, F.M.H. (2012). Effect of coating materials (gelatin) and storage time on internal quality of chicken and quail eggs under refrigerated storage. Egypt Poult Sci J. 32(1): 107-115.

28. Murray, M.W., Rutherford, P.P. (1963). The relationship between the loss of water and carbon dioxide from eggs and the effect upon albumen quality. Poult Sci. 42(2): 505-508. https://doi.org/10.3382/ps.0420505

29. Eke, M.O., Olaitan, N.I., Ochefu, J.H. (2013). Effect of storage conditions on the quality attributes of shell (table) eggs. Niger Food J. 31(2): 18-24. https://doi.org/10.1016/S0189-7241(15)30072-2

30. Collins, C.H., Lyne, P.M. (1976). Microbiological methods, 4th ed. London, UK: Butterworths

31. Şeremet, Ç. (2024). Protein, lipid and polysaccharide biomolecules in the coating of table eggs. Turk J Agric Food Sci Technol. 12(11): 1953-1958. https://doi.org/10.24925/turjaf.v12i11.1953-1958.6998

32. Silversides, F.G., Villeneuve, P. (1994). Is the Haugh unit correction for egg weight valid for eggs stored at room temperature? Poult. Sci. 73(1): 50-55. https://doi.org/10.3382/ps.0730050

33. Olawumi, S., Christiana, B. (2017). Phenotypic correlations between external and internal egg quality traits of Coturnix quails reared under intensive housing system. J Appl Life Sci Int. 12(3): 1-6. https://doi.org/10.9734/JALSI/2017/33802

34. Alig, B.N., Malheiros, R.D., Anderson, K.E. (2023). Evaluation of physical egg quality parameters of commercial brown laying hens housed in five production systems. Animals (Basel). 13(4): 716. https://doi.org/10.3390/ani13040716 PMid:36830504 PMCid:PMC9951926

35. Kumbár, V., Trnka, J., Nedomová, Š., Buchar, J. (2015). On the influence of storage duration on rheological properties of liquid egg products and response of eggs to impact loading-Japanese quail eggs. J Food Eng. 166, 86-94. https://doi.org/10.1016/j.jfoodeng.2015.05.030

Please cite this article in press as: Pekel A.Y., Yıldız A.E., Kızıl A., Aydın A. Influence of relative humidity on internal quality, bacterial load, and mold contamination of eggs during 28-day storage. Mac Vet Rev 2026; 49 (1): i-xi. https://doi.org/10.2478/macvetrev-2026-0022